

Staff Information

HSL Pathology Handbook

East and North Hertfordshire
NHS Trust



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Introduction

The Pathology service at East and North Hertfordshire NHS Trust (ENHT) is provided by Health Services Laboratories (HSL), for more information please see [Home | Health Services Laboratories](#).

Our aim is to develop and deliver a sustainable business that delivers quality and value for everyone we work with. We recognise that this must be achieved in a responsible manner and are committed to ensuring that our activities, carried out impartially, have a positive impact on both the communities in which we operate, and the wider healthcare sector and our services are delivered to patients free of discrimination.

To achieve this, we will:

- Play a key part in the development of healthcare in the UK. We will listen carefully to the requirements of NHS commissioners and the people they are commissioning for and develop quality based solutions that reflect their needs.
- Engage with staff to ensure that they are treated fairly, enable them to realise their full potential, and have an active role in developing the UK's clinical pathology workforce.
- Minimise any negative impact our business has on the environment. We have implemented a documented Environmental Management System based on the requirements of ISO 14001.

We offer a comprehensive range of pathology services. The Pathology service at Lister Hospital comprises a Blood Sciences Department (Clinical Biochemistry, Haematology & Blood Transfusion) and a Cellular Pathology Department (Histology and Diagnostic cytology). The Blood Sciences Laboratory offers a 24/7 service, and Cellular Pathology offer a weekday service.

Accreditation and Quality assurance

HSL is committed to providing doctors with pathology of the highest quality.

The quality of results is of fundamental importance, and HSL operates to stringent UK regulatory and International standards. Internal quality assurance is achieved by strict adherence to standard operating procedures for all analytical processes.

Quality assurance is administered by HSL's Quality Management Group (QMG). The QMG supports all HSL departments, to achieve and maintain the requirements of all relevant regulatory and accreditation standards. These include but are not limited to international standards such as ISO 15189, and UK regulations such as Blood Safety and Quality Regulations, and the Health and Social Care Act 2012. For an up-to-date list of accredited tests performed at Lister Hospital please see [UKAS 8846 Medical Multiple](#), other HSL laboratories carry their own Schedules of Accreditation available from the UKAS Website. HSL is a third party provider to the ENHT HTA licence (licence number 12110).

The team is primarily responsible for the implementation and management of a quality management framework, including document control and auditing processes, along with more technical elements associated with change management processes and validation framework. Our objective is to deliver a framework, that supports our services, with, high quality, safe and patient focussed service provision at its core.

Led by the director of governance, the team also includes quality advisors, quality managers, a quality administration team, and quality officers.

HSL participates in accredited External Quality Assessment Schemes. These schemes are subscribed to by NHS and private laboratories. Results are subjected to both internal and external quality control. Where no such scheme exists sample exchange schemes are carried out.

Details of the laboratories that HSL refers specialist testing to are available from HSL Referrals. These laboratories are UKAS accredited, or of equal accreditation status.

Data Protection

It is the policy of Health Services Laboratories (HSL) supported by its board of directors, to take steps to ensure that your information is kept confidential and secure and to otherwise protect and respect your privacy. HSL will only ever collect and process the minimum amount of information required in order to provide our pathology services. As well as the steps set out in this policy, HSL is accredited to the international standard for Information Security Management Systems set out in ISO/IEC 27001, our certificate may be found at [Sonic Healthcare Holding Company - IS 655966](#)

This is a high level privacy notice describing the information that HSL processes, the purpose of that processing, and how we protect it. For more detailed information including the lawful basis for processing please read the ['Detailed Privacy Notice'](#).

Data Controller

HSL is a part of The Doctors Laboratory Group, the largest independent provider of clinical laboratory diagnostic services in the UK providing pathology services to the private and public sector, information about the companies that comprise our group can be found at the respective websites: www.tdlpathology.com

www.hslpathology.com

This policy together with your terms and conditions sets out the basis on which any information HSL collects from you, or that you provide to HSL, will be processed. The following explains our views and practices regarding your information and how we treat it.

In providing products and services, HSL may be acting as a data processor on behalf of third parties (such as clinicians, hospitals and/or insurers) who will themselves be the data controllers, or as a data controller (if for example you are an employee). Where acting as a data controller, HSL will comply in full with this policy. Where acting as a data processor, HSL will be required to act on the instructions of the data controller.

Questions, comments and requests regarding this privacy policy are welcomed and should be made to:

HSL Data Protection
The Halo Building
1 Mabledon Place
London
WC1H 9AX

dataprotection@hslpathology.com

Key Contacts

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Cellular Pathology Manager

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Mohammed Syed

Sample Reception Manager

Mohammed.Syed@hslpathology.com

Abigail Toms

Quality Manager

Abigail.toms@hslpathology.com

| Department | Contact Number |
|---|----------------------------|
| Phlebotomy at Lister Hospital | ■ 01438 285229 |
| Phlebotomy at QEII Hospital | ■ 01707 247544 |
| Phlebotomy at Hertford County Hospital | ■ 01992 823200 |
| Lister Hospital Laboratory Blood Sciences Reception | ■ 01438 285461 |
| Lister Hospital Laboratory Biochemistry | ■ 01438 284690 |
| Lister Hospital Laboratory Haematology | ■ 01438 284961 |
| Lister Hospital Laboratory Blood Transfusion | ■ 01438 285245 |
| Lister Hospital Laboratory Cellular Pathology | ■ 01438 285198 |
| Lister Hospital Laboratory Out of Hours | ■ As above or use Alertive |
| Major Haemorrhage | ■ Ext 2222 |

Please phone your GP directly for your results. Do not phone the laboratory or Phlebotomy, as staff are not permitted to issue results.

Opening Hours

| Department | Hours |
|--|--|
| Lister Hospital ESL Core Opening Hours (Monday to Friday) | <ul style="list-style-type: none">■ 09:00 – 17:00 |
| Lister Hospital ESL Out of Hours Services (Blood Sciences) | <ul style="list-style-type: none">■ 17:00 – 09:00 (Weekdays)■ 24 Hours (Weekends and Public Holidays) |
| Phlebotomy at Lister Hospital | <ul style="list-style-type: none">■ 08:00 - 17:30 (Monday to Friday)■ Children under 10 walk in service 08:00 - 16:00■ 08:45-12:45 (Saturday) |
| Phlebotomy at QEII Hospital | <ul style="list-style-type: none">■ 08:00 – 17:30 (Monday to Thursday)■ 08:00 – 17:00 (Friday)■ Children under 10 by appointment only■ 08:00-12:00 (Saturday) |
| Phlebotomy at Hertford County Hospital | <ul style="list-style-type: none">■ 08:00 – 16:00 (Monday to Friday) |

Requesting Pathology Tests

The person making the request is responsible for ensuring the data on the request form is accurate, complete, and legible (whether hand-written or generated using ICE Order Communications System). Please give relevant clinical information to support all requests including relevant drug history, anatomical site of origin of specimen etc where relevant. Inadequately completed requests may lead to unnecessary delays, or rejection of samples.

The requester is also responsible for obtaining informed consent for the investigations; specific consent must be obtained if specimens may be used for purposes other than direct patient care (e.g. for research purposes).

The requester must raise the alert regarding patients with the same or similar names at their location by use of the "SAME NAME" sticker. Where needed, the requester must also identify the High-Risk status of the patient.

If using addressograph labels for patient details on duplicate/triplicate paper forms, these must be applied to each copy as the forms are often separated in the laboratory.

ICE requesting should always be used if available. It is not yet fully available for Blood Transfusion. To obtain user accounts and training for ICE, contact the ICE Team at enhtr.iceocs@nhs.net or phone Lister extension 4798.

For further information on the use of ICE, visit the Knowledge Centre>Pathology>ICE Order Comms.

The request form at the end of this handbook can be used to make manual requests when ICE is not available. If using addressograph labels for patient details on manual requests this must be applied to the form.

Sample identification and acceptance criteria

Requests that do not meet the acceptance criteria will be rejected.

Correct use of ICE fulfils all the minimum data set requirements for request forms:

- NHS number (or CHI [Scotland] or Health and Care Number [Northern Ireland])
- Full name (surname and forename)
- Date of birth and/or hospital number
- Gender
- Location and destination for report
- Responsible Consultant/ GP/ Dentist
- Investigations required must be clearly indicated

Useful information to enable appropriate analysis, interpretation and reporting:

- Name of the requester
- Contact number of the requester
- Relevant clinical information
- Other test-dependent required information (eg. time of last dose of certain drugs, anatomical site of sample, recent foreign travel where relevant etc)
- Date and time of sample (24 hour clock)
- Name/signature of person collecting the sample

For patients without NHS numbers, or those who require anonymisation:

The responsible Consultant is responsible for accepting and correctly assigning costs for non-NHS patients (eg. overseas visitors). The laboratory will accept samples without NHS number on this basis, as long as full name, date of birth and either our local hospital number or the first line of the address is supplied.

Patients of Genito-Urinary Medicine, staff requiring investigations by Health at Work, and (rarely) some participants in research trials may justifiably be anonymised. In these cases, NHS numbers are not supplied. The laboratory will accept these samples as long as a unique coded identifier is supplied.

This takes the place of both the NHS number and the patient's full name on the sample and request form.

For unidentified patients associated with Major Incidents:

Due to the increased potential for confusion and error, all samples from unidentified patients during Major Incidents must be labelled with a unique identification number. These are included in the AE packs ready for major incident. Please see below;

NHS No:
Patient ID: RWH3022539
Name: MAJOR INCIDENT 001, Patient
001,
Address: No fixed abode, No fixed
abode, No fixed abode, No fixed abode,
ZZ99 3CZ
DOB: 02/02/1900

For Temporary Identification of the Unknown Patient follow the Trusts Policy CP274 where randomised names based on an amended version of the phonetic alphabet are used. An estimated date of birth is used and if the sex of the patient is unknown then they are set to female due to potential RhD-ve status and risk of sensitisation if RhD +ve blood is administered.

For Private patients and Category II requests:

All Private and Category II requests must be declared as such on the request form.

If no NHS number or other unique coded identifier is provided, the first line of the patient's address must be supplied.

The requester is responsible for ensuring that the patient has already signed an "agreement to pay" form. If a request is suspected to be non-NHS there may be a delay in testing until payment is secured.

Specimens submitted from private patients are investigated by the same protocols as NHS patients.

Private Patients attending phlebotomy with requests from other hospitals will be asked to sign an 'Agreement to pay' form before being bled.

Please contact the Trust's Private Patients Team if you require further information, such as prices.

Collecting Pathology Specimens

For conscious patients, the person obtaining a specimen must positively identify them by asking their surname, first name and date of birth, and checking that these match the request form. For inpatients, these details should be further checked with those on the patient's wristband (which will also have NHS and hospital numbers). For specimen collection from outpatient areas where patients do not have wristbands, the patients should be asked to also recite the first line of their address for checking against the request form.

If the patient is unconscious, a senior member of the ward staff should confirm the patient's details in addition to the wristband check.

The same person who collected the specimen must themselves label the container correctly and fully in the presence of the patient immediately after collection. Do not hand samples to a colleague for labelling. Do not pre-label blood tubes.

Minimum data set requirements for specimen containers:

- NHS number (or CHI [Scotland] or Health and Care Number [Northern Ireland])
- Full name (surname and forename)
- Date of birth and/or hospital number
- Specimens for cross-matching must always be signed by the person obtaining the specimen
- When several samples from the same patient are to be collected they must all have the correct corresponding sample label from the appropriate request form. Ie F barcode number for fluoride oxalate sample for Gluc. The ICE request form gives this information at the bottom.

As well as using the above identity information, all specimen labels should include the following:

- date and time of specimen collection
- initials of person collecting the sample

There is zero tolerance for samples and request forms for Blood Transfusion that are not labelled with the NHS number (where a patient has one). Samples and request forms without this information will not be processed unless the patient requires urgent blood product support.

Where no NHS number is available, see the requirements listed above under "Requesting pathology tests".

Addressograph or ICE labels may be used for the majority of samples but **must not** be used for blood transfusion samples.

Unlabelled or mislabelled specimens cannot be accepted by the laboratory for testing. Specimens that cannot be repeated (e.g. CSF, biopsies etc) may be accepted after full discussion with a senior member of laboratory staff, and logging of an Enhance incident by the person responsible.

Tips for obtaining valid results and to avoid sample rejection

The laboratory endeavours to provide the best possible service but the quality of the results ultimately depends on the quality of the sample submitted for testing

- Avoid prolonged venous stasis when collecting blood.
- Use the correct order of draw - consult the tube guide for order of drawing samples.
- Avoid contamination of sample with iv fluids (do not use drip arm).
- Do not mix blood from one specimen container with another.
- Ensure that urine collections are timed correctly and kept cool.
- Send samples to the laboratory or specimen collection points without delay.
- Be aware of sample fill levels especially for coagulation samples, ESR and Quantiferon-TB. Always fill to/ within the black fill mark on the side of the tubes to avoid sample rejection.
- **DO NOT** send blood gas samples via the chute.
- Always check sample tube expiry dates. Out of date tubes will be rejected.
- Ensure all samples sent are securely sealed, leaking samples will not be processed.
- Avoid storing samples in direct sunlight or on/near radiators
- Do not store uncentrifuged biochemistry samples in a fridge, samples should be kept as close to 22 degrees Celsius as possible.
- Courier samples as soon as possible to the laboratory

Transport of specimens to the Pathology Department

All specimens must be transported inside a fully sealed polythene specimen bag. Place only one patient's form and its associated specimens into one bag. Place the specimen in the welted pouch and seal it, place the form into the adjoining unwelted pouch (for blood transfusion samples the bag is integral to the request form). Do not place the form in the welted pouch with the specimen.

Specimens should be sent to the laboratory by either the pneumatic chute or by a hospital porter. High-risk samples, blood gas samples requiring PO_2 and PCO_2 measurement, blood culture samples, Microbiology swabs, and Cellular Pathology samples must not be sent through the pneumatic tube system.

Blood gas syringes must be sealed with the caps supplied. **Never send syringes with needles still attached.** They must be transported to the laboratory as soon as possible and within 20 minutes. Syringes received via the chute can only be tested for pH, electrolytes, glucose, lactate.

Specimens from Queen Elizabeth II and Hertford County Hospitals are collected in Phlebotomy clinics then transported to Lister Pathology departments. Specimens referred for testing elsewhere may be transported in different ways, including Royal Mail, Hays DX, NHTSBT, TDL/HSL employed drivers. Additional third party couriers may be used if required.

Urgent Samples

Samples from agreed priority areas (e.g. ED) or clinically urgent samples from other areas should be placed in a red welted pouch and sent immediately to the lab for processing. Urgent samples from non-priority areas of the hospital must be heralded to the laboratory by phoning 5461.

General Practitioner practices

Samples are collected from Surgeries and other phlebotomy locations by Couriers and delivered to Lister Hospital or other HSL sites as appropriate. Individual welted specimen bags are first packed into a large bag as outlined in the table below.

| Test Request | Bag | LIMS | Drop Location | Location of Analysis | Discipline |
|----------------------------|---------|----------------------|---------------|---|---|
| Routine GP Requests | ■ Clear | ■ Helix | ■ Barnet | ■ Barnet>Halo ■ Barnet>RFL (Special coagulation) | ■ Haematology (Barnet) ■ Biochemistry (Halo) ■ Immunology (Halo) ■ Microbiology (Halo) |
| Urgent GP Requests | ■ Red | ■ Helix | ■ Barnet | ■ Barnet>Halo ■ Barnet>RFL (Special coagulation) | ■ Haematology (Barnet) ■ Biochemistry (Halo) ■ Immunology (Halo) ■ Microbiology (Halo) |
| Routine Hospital Requests | ■ Green | ■ Helix ■ WinPath | ■ Lister | ■ Lister ■ Halo ■ Referral Lab | ■ Haematology (Lister) ■ Biochemistry (Lister or Halo) ■ Immunology (Halo) ■ Microbiology (Halo) |
| Urgent Hospital Requests | ■ Red | ■ Helix ■ WinPath | ■ Lister | ■ Lister ■ Halo ■ RFL ■ Referral Lab | ■ Haematology (Lister) ■ Biochemistry (Lister or Halo) ■ Immunology (Halo) ■ Microbiology (Halo) |
| Hospital Blood Transfusion | ■ Green | ■ WinPath | ■ Lister | ■ Lister | ■ Blood Transfusion |

| | | | | | |
|--------------------------------------|--|--|---|--|---|
| GP Antenatal Requests | <ul style="list-style-type: none">■ Green | <ul style="list-style-type: none">■ Helix■ WinPath (BT) | <ul style="list-style-type: none">■ Barnet■ Barnet>Lister | <ul style="list-style-type: none">■ Lister■ Halo (confirmation)■ Lister (BT) | <ul style="list-style-type: none">■ Blood Transfusion■ Everything Else |
| Histopathology Hospital Requests | <ul style="list-style-type: none">■ Green | <ul style="list-style-type: none">■ WinPath | <ul style="list-style-type: none">■ Lister | <ul style="list-style-type: none">■ Lister | <ul style="list-style-type: none">■ Histopathology |
| Histopathology GP Requests | <ul style="list-style-type: none">■ Green | <ul style="list-style-type: none">■ WinPath | <ul style="list-style-type: none">■ Barnet>Lister | <ul style="list-style-type: none">■ Lister | <ul style="list-style-type: none">■ Histopathology |
| GP Cervical Screening Requests | <ul style="list-style-type: none">■ Purple | <ul style="list-style-type: none">■ N/A | <ul style="list-style-type: none">■ Barnet>Lister | <ul style="list-style-type: none">■ NNUH | <ul style="list-style-type: none">■ Gynae Cytology |
| Hospital Cervical Screening Requests | <ul style="list-style-type: none">■ Purple | <ul style="list-style-type: none">■ N/A | <ul style="list-style-type: none">■ Lister | <ul style="list-style-type: none">■ NNUH | <ul style="list-style-type: none">■ Gynae Cytology |

FOR REFERENCE ONLY

High Risk Specimens

Separate procedures are used in the laboratory for the safe handling and examination of samples from patients known or suspected to have infections caused by high risk (hazard group 3 & 4 pathogens - see below) that pose a risk to laboratory workers and others if handled incorrectly. Under the Health & Safety at Work etc Act 1974 it is the responsibility of the person making pathology requests to ensure that the laboratory is appropriately informed of the potential dangers of handling any high risk samples.

Requests made via ICE should be flagged by ticking the high risk box in the order entry screen on ICE. Paper/ICE requests must give sufficient clinical information to enable experienced laboratory staff to know what special precautions are necessary, including relevant foreign travel history.

In the interests of confidentiality 'High Risk' labels should be placed only on the sample.

Hazard Group 3 & 4 Pathogens

| Pathogen | Hazard group | Pathogen | Hazard group |
|-------------------|----------------------------|-------------------------------------|----------------------------|
| Alpha viruses | <input type="checkbox"/> 3 | Herpesvirus simiae | <input type="checkbox"/> 4 |
| Blastomycetes | <input type="checkbox"/> 3 | HTLV 1 and 2 | <input type="checkbox"/> 3 |
| Brucella sp. | <input type="checkbox"/> 3 | Leishmania | <input type="checkbox"/> 3 |
| Borna Virus | <input type="checkbox"/> 3 | Monkeypox | <input type="checkbox"/> 3 |
| Claophialophora | <input type="checkbox"/> 3 | Mycobacterium sp | <input type="checkbox"/> 3 |
| Coccidioides | <input type="checkbox"/> 3 | Naegleria | <input type="checkbox"/> 3 |
| Covid-19 | <input type="checkbox"/> 3 | Nipah | <input type="checkbox"/> 4 |
| Duvenhage | <input type="checkbox"/> 3 | Paracoccidioides | <input type="checkbox"/> 3 |
| Ebola viruses | <input type="checkbox"/> 4 | Penicillium marneffei | <input type="checkbox"/> 3 |
| E.coli O-157 | <input type="checkbox"/> 3 | Phleboviruses | <input type="checkbox"/> 3 |
| Flaviviruses | <input type="checkbox"/> 3 | Piry | <input type="checkbox"/> 3 |
| Hanta viruses | <input type="checkbox"/> 3 | Plasmodium falciparum | <input type="checkbox"/> 3 |
| Histoplasma | <input type="checkbox"/> 3 | Rickettsia sp | <input type="checkbox"/> 3 |
| HIV 1 and 2 | <input type="checkbox"/> 3 | Salmonella paratyphi | <input type="checkbox"/> 3 |
| Hepatitis B virus | <input type="checkbox"/> 3 | Salmonella typhi | <input type="checkbox"/> 3 |
| Hepatitis C virus | <input type="checkbox"/> 3 | Simian Immunodeficiency virus (SIV) | <input type="checkbox"/> 3 |
| Hepatitis D virus | <input type="checkbox"/> 3 | Trypanosoma | <input type="checkbox"/> 3 |
| Hepatitis E virus | <input type="checkbox"/> 3 | Variola (minor & major) | <input type="checkbox"/> 4 |
| Hepatitis G virus | <input type="checkbox"/> 3 | | <input type="checkbox"/> |

Also the causative agents of:

| Agent | Hazard group | Agent | Hazard group |
|--------------------------------------|----------------------------|-------------------------|----------------------------|
| Anthrax | <input type="checkbox"/> 3 | Plague | <input type="checkbox"/> 3 |
| Bovine spongiform Encephalitis (BSE) | <input type="checkbox"/> 3 | Pork tape worm | <input type="checkbox"/> 3 |
| Brucellosis | <input type="checkbox"/> 3 | Q-fever | <input type="checkbox"/> 3 |
| Creutzfeldt-Jakob disease (CJD) | <input type="checkbox"/> 3 | Rabies | <input type="checkbox"/> 3 |
| Dengue | <input type="checkbox"/> 3 | SARS | <input type="checkbox"/> 4 |
| Diphtheria | <input type="checkbox"/> 3 | Smallpox | <input type="checkbox"/> 4 |
| Dog tapeworm | <input type="checkbox"/> 3 | Tick borne encephalitis | <input type="checkbox"/> |

| | | | |
|-------------------------|-----|---|-----|
| Dysentery | ■ 3 | Transmissible spongiform encephalopathies (TSE) | ■ 3 |
| Fatal familial insomnia | ■ 3 | Tularaemia | ■ 3 |
| Glanders | ■ 3 | Typhoid | ■ 3 |
| Hendra | ■ 4 | Variant Creutzfeldt-Jakob disease (vCJD) | ■ 3 |
| Kuru | ■ 3 | Viral Haemorrhagic Fever (Lassa Fever, Ebola Fever and Marburg Disease) | ■ 4 |
| Melioidosis | ■ 3 | Yellow Fever | ■ 3 |
| Paratyphoid | ■ 3 | | ■ |

Covid-19

This is a Hazard Group 3 organism. There is evidence that blood contains very low viral load, hence blood samples may be sent to the laboratory without a High Risk label and sent using the pneumatic chute. Respiratory samples including viral swabs and nasopharyngeal aspirates or sputum samples have high viral load hence are high risk and must not be sent via the chute.

Reporting Results

Access to completed pathology results is available throughout the Trust sites (e.g. wards and departments), CCG sites (e.g. health centres, GP surgeries), Hertfordshire Partnership units, and to other customers via ICE OCS (order communications system). If you are accessing results through East & North Hertfordshire NHS Trust ICE OCS then assume that all tests are processed at an HSL site, either Lister, Halo or Barnet (for GP samples).

Critical results (for details see individual discipline sections below) are telephoned urgently to the requesting clinician, ward or clinical team. Please ensure that correct information is on all requests otherwise delay is likely. The receiving clinician (who must be a qualified member of nursing, midwifery or medical staff; or a Clinical Support Worker who has undertaken specific training) should document the results within the patients notes at the earliest opportunity and bring it to the attention of a senior member of staff where necessary. In line with the Trust's policy 'Acting on Diagnostic Investigations' it is the requesting clinician's responsibility to review all pathology results from tests requested and to act on these results appropriately.

Requesting further tests on samples already in the laboratory (Add-tests)

The laboratory will respond to add-test requests as quickly as possible, but this process is inefficient and slow in comparison with the usual process of receiving samples with the request, so we ask requesters to please try to avoid the need for these as far as possible. Although the sample may already be received in the laboratory it may not be possible to retrieve it from our automated systems for a considerable length of time. If the request is very urgent, it may be quicker to re-bleed the patient and send a fresh specimen.

If an add-test is required, please make a new ICE request for the additional test(s), citing in the "clinical details" section the words "add test" and the laboratory number of the sample to which you need to add further tests:

To obtain the laboratory number for a sample, look in "view requests by patient", click on "view order" and it is the number identified as "order accession number" and follows the format of YYL123456 where YY is the year.

To obtain the laboratory number for a sample that has already been reported, it is also possible to obtain it by looking in "view patient reports", where it is the number appearing in the "sample number" column on the screen.

Not all add-test requests can be accommodated, due to sample stability or storage issues.

In the event of major delays to the system the Pathology department will notify the matron/bed manager on call for the hospital. In more severe circumstances or when planned downtime is known of in advance, notification will be made via communication via email with the Trust.

Routinely if any examination is delayed that could compromise patient care, the requestor will be notified.

Pathology Information

Pathology Supplies

Following the change in Pathology Service provider across the Herts & West Essex (HWE) Integrated Care System, to be provided by Health Services Laboratories (HSL), there will be changes to the way pathology supplies are ordered by East & North Herts Primary Care and Community Services. These new processes will begin on 1st April 2025.

- HSL have created a single order form. To access this please use the following link:
https://pathologyforms.formstack.com/workflows/enhgp_pathology_supplies_order_forms1
- Ensure your correct practice and location is selected from the first drop down list.
- Complete your requester details (name / email address / contact phone number).
- Input the item quantity in the required field allocated to each product. If there is a limit on how many can be ordered, this will be indicated in small print next to the product description. For items you do not require, leave them blank.
- Once complete, please select the submit button.
- A confirmation email will be sent to the email address provided within the form.
- The form will be received by our Supplies team, who will process your order and arrange for this to be delivered to you.
- For any queries or changes regarding an order you can contact the Supplies team on:
 - ls.helpdesk@hslpathology.com
 - 020 7307 9440 quoting the order reference number.

If your request is urgent and you need your order to be expedited for urgent delivery, please submit the online form and follow-up with a phone

Point of care testing

HSL are responsible for providing advice and support for Point of Care Testing (POCT) for Blood Gas and Full Blood Count. Tests must only be performed by trained and competency assessed users. Please contact HSL POCT with any queries about maintenance, quality control, training, audits, support issues, operation problems or Health and Safety aspects. The East and North Hertfordshire NHS Trust Policy on POCT must be followed. Before purchase of POCT devices, the Trust POCT Committee must be consulted about suitability.

Other services and complaints

Pathology can provide a range of services and information to wards, departments and GP practices. If you wish to discuss any service developments or require information relating to or derived from the pathology service then please direct your enquiry as follows:

To make a complaint email: complaintsadmin.enh-tr@nhs.net for Trust complaints or

hwe.complaints@tdlpathology.com for laboratory complaints.

To log an incident :- Trust users report on the Trust Enhance system

GP's go through the GP Liaison team – Gpliaison.enh-tr@nhs.net

Phlebotomy

The Phlebotomy department is part of HSL and supports phlebotomy clinics across sites at Lister Hospital Stevenage, The New QEII Welwyn Garden City and Hertford County Hospital.

The phlebotomy service supports GP surgeries in the area and also offers a local domiciliary service.

Responsibility for ensuring consent for pathology tests lies with the requester. Consent to blood tests is assumed when a patient attends the phlebotomy department with a request. For some specialist tests and research, separate consent with the requesting clinician will be sought.

Please note that remnant samples (after all requested tests have been completed) may be anonymised and used for service evaluation or quality assurance purposes.

Key contacts

HSL HWE Phlebotomy manager

Elaine Stokes

elaine.stokes@nhs.net

ENHT Phlebotomy manager

Debra Thomas

debrathomas@nhs.net

GP patient Blood tests and online booking

Blood test appointments, requested by GPs, should now be booked online for the following hospitals: Hertford County Hospital, Lister Hospital and New QEII. This is to help maximise the number of tests we can offer to patients, minimise queues.

Our current walk-in service will remain in place for those attending outpatient appointments, patients with an urgent GP blood request and those with a request for specialised tests, for example where a blood test is required on a certain day of the menstrual cycle.

Adults and children (11 years and over)

Book appointments using the links below:

[Lister Hospital](#)

[New QEII Hospital](#)

[Hertford County Hospital](#)

Children (10 years and under)

If the patient is under 10 years old, please book using the following link:

[Lister Hospital \(Children\)](#)

There are limited appointments for children under 10 available at Hertford County Hospital and New QEII Hospital. Please call the Phlebotomy team below to book.

Appointment booking lines 01438 284044/ 01438 284330

Emla cream (a topical numbing cream) can be obtained by prescription from the GP or bought over the counter at any pharmacy. It needs to be applied 1 hour before attending the department. This may be useful for use in children.

Numbing spray is also available for children and needle phobic adults. This is applied immediately prior to venepuncture.

Attending your appointment:

When you arrive for your appointment, please go to the phlebotomy reception area at the hospital and provide your name and your appointment time.

Please arrive promptly and ideally no earlier than five minutes beforehand to help ensure social distancing.

What if I am unable to book online?

If you are unable to book online, please call the phlebotomy team on 01438 284044 / 01438 284330 Monday-Friday between 9-5pm. The team will be happy to book your appointment for you.

Please note: In order to book your appointment, you will need your:

- Name
- Date of birth
- Postcode
- Mobile phone number
- NHS number – you can easily find out your NHS number on the NHS website

Please be advised these lines can get extremely busy so if you have someone to help you access the online booking system this may be quicker.

Appointments are needed to attend Phlebotomy at:

Baldock

Park Drive Health Centre (by appointment only via Baldock and Letchworth GP surgeries)

Tuesday, 8.30am to 12noon

Wednesday, 1.30pm to 4.30pm

Thursday, 8.30am to 12noon

Hitchin

Bedford Road Health Centre (by appointment only and then through just the following Hitchin GP surgeries: Marshall House, Orford Lodge, Portmill and Regal Chambers)

Tuesday, 8am to 4.30pm

Hoddesdon

Hoddesdon Health Centre (by appointment only)

Monday to Friday, 8.30am to 2pm (excluding bank holidays)

Stevenage

Danestrete Clinic (by appointment only – appointments are made normally directly by nurses with patients attending anti-coagulation clinics)

Mondays and Wednesdays only, 8.30am to 11.45am (excluding bank holidays)

Waltham Cross

Stanhope Road Health Centre (by appointment only, but through the health centre itself – 01992 818500)

Monday to Friday, 8.30am to 12noon (excluding bank holidays)

Ware

Bowling Road Health Centre (by appointment only)
Monday, 1.30pm to 3.45pm (excluding bank holidays)
Wednesday, 8.30pm to 12noon
Friday, 8.30am to 12noon (excluding bank holidays)

Fasting blood test requirements

Fasting glucose – at least 8 hours fasting
Glucose tolerance test (GTT) – 8-14 hrs fasting
Fasting lipids and triglycerides – at least 12-14 hrs fasting
NB: Fasting means no food or drink apart from water.

24 hour urine samples

If you have been asked to complete a 24 hour urine collection the containers can be picked up from the Phlebotomy receptions at QE2, Hertford and Lister Hospitals. Information leaflets are available for advice on how to fill.

Some tests require a container that is issued with a small amount of acid to preserve the sample. Do not discard; this must remain in the bottle. Completed samples should be returned to any phlebotomy reception.

Stay hydrated

It is advisable to be well hydrated before a blood test. Please remember to drink water before getting your blood taken.

Inpatient blood tests

All ICE requests for ward rounds need to be made and on the ICE system by 7am on the day of collection.
Phlebotomists start bleeding from 6am to 11am (7am – 11 at weekends) however priority wards are visited first.
Please be aware that not all tests are suitable to be collected by phlebotomists on ward rounds. These include but are not limited to

- blood cultures
- timed drug/ therapeutic monitoring samples
- Dynamic function tests
- Time sensitive tests ie Ammonia, ACTH, Insulin etc.

Please **DO NOT** request these tests for ward rounds.

Domiciliary blood tests

This service is highly oversubscribed. It is only offered to patients who are completely housebound or are severely unwell.
Patients meet the criteria for a domiciliary visit if they are;

- Housebound
- On oxygen
- In a care/nursing home
- Visited by their GP at home (this is at the discretion of the GP)
- Extremely unwell patients (this is at the discretion of the requesting clinician)

Requests for this service on patients must be emailed to:

Listerdomiciliary.enh-tr@nhs.net, Hertforddomiciliary.enh-tr@nhs.net, QE2domiciliary.enh-tr@nhs.net

Patient Results

Please phone your GP directly for your results.

Do not phone the laboratory or Phlebotomy, as staff are not permitted to issue results.

Clinical Biochemistry

Clinical Biochemistry is part of Blood Sciences department at ENHT, this information is related to all acute work performed with ENHT

Opening hours

Monday to Friday: 09:00-17:00 At all other times contact the Biomedical Scientist on call for urgent work only.

General queries: 01438 284690

Key contacts

Consultant chemical pathologist

Dr Adie Viljoen

E: adie.viljoen@nhs.net

T: 01438 285972 (Ext 5972)

Consultant biochemist and point of care committee chair

Mrs Angela Woods

E: angelawoods@nhs.net

T: 01438 286145 (Ext 6145)

Essential Services Laboratory Manager

Kate Barrett

E: Kate.barrett@hslpathology.com

Point of Care Testing co-ordinator

Julia Dowsett

E: Julia.dowsett@nhs.net

T: 01438 288018 (Ext 8018)

Point of Care Testing Associate Practitioner

Bhranti Vashi

E: bhranti.vashi@hslpathology.com

Results

Pathology results are available on Sunquest ICE system, if results are not available please contact the laboratory on Ext 5461

Limitations of biochemical test results

Please be aware that biochemistry tests are generally robust but all are vulnerable to occasional assay interference due to uncommon patient-specific factors. Clinicians should always correlate results with the clinical picture. If a patient's results do not match their clinical condition, please discuss with one of the laboratory consultants so that further testing can be arranged if needed.

Special note regarding immunoassay tests

All immunoassay tests (troponin, hormones, haematinics, tumour markers etc) have the potential to produce erroneous results in patients with unusual antibodies.

In recent years, dietary supplements containing high-dose biotin (up to 650 times the recommended daily intake) have been marketed for supposed skin, hair and nail benefits. Even higher doses of biotin are also starting to be used in progressive multiple sclerosis. High dose supplementation ($\geq 5\text{mg/day}$) can interfere with all immunoassay tests that use biotin/streptavidin in their design, this includes all those performed in the Lister Hospital Laboratory. The expected pattern of results with high-dose biotin interference is:

- falsely high thyroid hormones, steroid hormones, digoxin
- falsely low peptide and protein hormones (TSH, troponin, PSA, PTH etc)

Severe interference from biotin can lead to biochemically thyrotoxic results that do not match the clinical picture.

There is lack of certainty about the time required to clear high-dose biotin from the blood, however between 1 and 4 days abstinence is probably advisable, depending on the magnitude of the dose and renal function.

Tests and specimens

Results are reported with reference ranges and/or an interpretative comment. Advice on appropriate requesting and interpretation of results is available at all times from the consultant staff by telephone, mobile telephone. Out of core hours the consultant on-call should be accessed through switchboard.

Add on requests will only be accepted for tests performed at Lister Hospital, these must be made within 24hours and must be accompanied by a request form.

Dynamic function tests – please discuss with the consultant chemical pathologist or the consultant biochemist.

Critical results

For Lister Hospital requests results outside the limits below will be telephoned urgently to the requesting clinician.

| Test | Units | Critical Low | Critical High |
|---|--------|------------------------------|----------------------------|
| Acute kidney injury (AKI) warning stage | | | 3 |
| Albumin | g/L | <15 | |
| ALT (paeds) on first occurrence | U/L | | >500 |
| Amylase | U/L | | 500 |
| Ammonia | µmol/L | | 100 |
| Bilirubin (newborns only) | µmol/L | | 250 |
| Calcium (adjusted) | mmol/L | 1.8 | 3.5 |
| Carbamazepine | mg/L | | 25 |
| Creatine kinase (CK) | U/L | | 5000 |
| Creatinine | µmol/L | | 500 |
| Cortisol (note below) | nmol/L | <100 (<18 yrs old only) | |
| Digoxin | ng/L | | 2.5 |
| FT4 | pmol/L | 6 | 40 |
| Gentamicin pre-dose (adults) | mg/L | | 1.0 |
| Gentamicin pre-dose (newborns) | mg/L | | 2.0 |
| Gentamicin post-dose | mg/L | | 10.0 |
| Glucose (random or fasting) | mmol/L | ≤2.5 | ≥15 (<16yr) ≥25 (>16yr) |
| Iron | µmol/L | | 55 |
| Lithium | mmol/L | | 1.0 |
| Magnesium | mmol/L | 0.40 | 2.00 |
| Paracetamol | mg/L | | 100 |
| Phenytoin | mg/L | | 25 |
| Phosphate | mmol/L | 0.40 | 3.00 |
| Potassium | mmol/L | 2.5 | 6.5 |
| Salicylate | mg/L | | 350 |
| Sodium | mmol/L | ≤130 (<16yr) ≤120 (>16yr) | 155 |
| Theophylline | mg/L | | 25 |
| Triglycerides | mmol/L | | 20.0 |
| Urate | µmol/L | | 1000 |
| Urea | mmol/L | | 30.0 |

Biochemistry blood tests

If the assay required is **NOT** listed then please contact the department directly. All results are reported with reference ranges and/or an interpretative comment. Advice on appropriate requesting and interpretation of results is available at all times from the Chemical Pathologists and Clinical Biochemists.

Sample volumes: Most routine tests can be performed on 5ml clotted (YELLOW) serum sample, unless otherwise stated below (For paediatric samples a minimum of 1ml of blood is required).

Indicative turnaround times are given for non-urgent situations. It may be possible to expedite results by discussing the clinical situation with the laboratory consultant staff.

If a required test is not listed here, please discuss with the laboratory consultant staff.

Please note that all reference ranges provided here are accurate for the Lister Hospital laboratory at time of publication, reference ranges from other laboratories testing samples may differ. All reports will quote a reference range related which must be used when reviewing results regardless of what is quoted here.

| Test | Sample | Laboratory Adult Reference range | Referred site code | Turnaround time | Comments |
|--|------------------------------------|--|--------------------|-----------------|---|
| 5-hydroxyindole acetic acid (5-HIAA) | Serum (Yellow) | <140nmol/L | MWY | <2 weeks | Collect sample after overnight fast, to avoid false positives from serotonin-rich foods. Do not eat walnuts in the 24h period prior to the blood collection. |
| This serum test replaces 24 hour urine 5-HIAA for the diagnosis of carcinoid syndrome, in which it is usually strikingly elevated. It is also used for monitoring neuroendocrine tumours (NETs) that have been shown to secrete 5-HIAA. For non-functional NETs consider using chromogranin A instead. False positive results may occur if the diet contains serotonin-rich foods (eg. banana, tomato, walnut, aubergine, avocado, pineapple, kiwi fruit). An overnight fast is sufficient to prevent this; however walnuts should not be consumed during the previous day as they are particularly rich in serotonin. 5-HIAA is eliminated by the kidneys, hence serum levels increase with decreasing GFR, and urine levels decrease. If GFR is below 60mL/min, it is likely that serum 5-HIAA will be elevated in otherwise normal individuals with no carcinoid tumour or NET. | | | | | |
| Acylcarnitines | 4 dried blood spots (Guthrie card) | Interpretation with report | Halo | <2 weeks | Clearly write "acyl carnitines" on the card. Allow to air-dry, then place in glassine sleeve and send to Laboratory. Do not post to Newborn Screening Laboratory. |
| The acylcarnitine test (bloodspot) is a core metabolic test useful in the investigation of fatty acid oxidation defects and classical organic acidurias. Investigations should be performed, where possible, when the patient is symptomatic. | | | | | |
| ACTH - Adrenocorticotrophin | EDTA (Lavender) | Interpretation with report | HALO | 2 weeks | Send to lab promptly for immediate freezing of plasma. Take blood for cortisol also. |
| Adrenaline [part of Plasma catecholamines] | Lithium Heparin (Green) | This test is not routinely offered. It is not a sensitive test for phaeochromocytoma – send plasma metadrenalines instead. | | | |
| Albumin | Serum (Yellow) | 35-50g/L | | <1 day | |
| Alcohol (ethanol) | Fluoride EDTA (Grey) | | | <1 day | Clinical samples only. Medico-legal work NOT accepted |

| | | | | | |
|---|-----------------|---|------|-------------|---|
| Aldosterone | EDTA (Lavender) | See comments | ADI | 2 weeks | Renin sample should be taken at same time. 90–405pmol/L (adult recumbent overnight) 90–720 pmol/L (random or upright) |
| Aldosterone test is used along with renin in the investigation of suspected primary hyperaldosteronism (Conn's syndrome) in patients with moderate/severe hypertension that is of early onset, drug-resistant, or associated with hypokalaemia or adrenal incidentaloma. See ARR below. | | | | | |
| Aldosterone /renin ratio (ARR) | EDTA (Lavender) | <91 pmol/mU | | | |
| ARR is the first-line investigation for suspected primary hyperaldosteronism. Before doing this test, try to correct hypokalaemia and encourage liberal sodium intake. If possible, withdraw the following drugs and products for at least 4 weeks as these have a big effect on ARR: spironolactone, amiloride, eplerenone, triamterene, potassium-wasting diuretics, liquorice. Many other drugs have lesser but predictable effects on ARR and drug history should always be reviewed when interpreting results. | | | | | |
| Blood samples should be collected in the morning after the patient has been up for at least 2 hours and seated for 5-15 minutes. | | | | | |
| High aldosterone-renin ratio is not diagnostic and should be further investigated. | | | | | |
| ALP (alkaline phosphatase) | Serum (Yellow) | 30-130 IU/L | | <1 day | Age- and sex-related reference ranges are on paediatric reports. |
| As of 4/2/2021, results for alkaline phosphatase (ALP) are increased by about 7% in order to improve alignment to international standards. Reference ranges remain unchanged as these are defined nationally for this test. We do not expect this small change to have any significant impact on clinical care. | | | | | |
| ALP isoenzymes | Serum (Yellow) | Interpretation with report | | 4 weeks | Rarely needed. Only done if ALP is markedly elevated |
| ALP Bone Specific [bone ALP] | Serum (Yellow) | Interpretation with report | | 3 weeks | |
| Alpha 1 antitrypsin | Serum (Yellow) | 1.1-2.1g/L 1w (0.9-2.2) 6m (0.8-1.8) 1yr (1.1-2.0) 5yr (1.1-2.2) 10yr (1.4-2.3) 15yr (1.2-2.0) (1.1-2.1) | HALO | <1 week | Ranges vary for children |
| Alpha 1 antitrypsin phenotype | Serum (Yellow) | Interpretation with report | ADI | 2 – 3 weeks | |
| AFP – alpha fetoprotein in pregnancy | | Interpretation with report | | | Arranged via maternity services |
| AFP – alpha fetoprotein (tumour marker) | Serum (Yellow) | 0-9 kU/L | HALO | <3 days | |
| Alpha subunit of pituitary hormones | Serum (Yellow) | Interpretation with report | QEH | <2 weeks | By prior arrangement with consultant only |
| ALT (alanine amino-transferase) | Serum (Yellow) | 7-40 IU/L | | <1 day | |

| | | | | | |
|--|-------------------------|---|------|-----------|---|
| Aluminium | Trace metal (Dark Blue) | 0-0.3µmol/L | Halo | <2 weeks | |
| Amino acids | Lithium Heparin (Green) | Interpretation with report | GOS | < 2 weeks | Send to lab promptly for immediate freezing of plasma. |
| Amino acids (plasma) are a core metabolic test useful for the investigation of urea cycle defects and some organic acidurias. Investigations should be performed, where possible, when the patient is symptomatic. | | | | | |
| Amiodarone | Serum without gel (Red) | 0.5-2.0mg/L | Halo | 1 week | By prior arrangement with consultant only |
| Amisulpride | EDTA (Lavender) | A target range of 100-400 µg/L plasma amisulpride (pre-dose) has been suggested at therapeutically effective doses from steady-state pharmacokinetic studies. | Halo | 2 weeks | By prior arrangement with consultant only |
| This test is occasionally needed for dose adjustment in patients prescribed anti-psychotics, who are on relevant co-medication, smoker etc or who are prescribed off-licence doses | | | | | |
| Amitriptyline | EDTA (Lavender) | Interpretation with report | | 1 week | By prior arrangement with consultant only |
| Ammonia | EDTA (Lavender) | 16-60umol/L 4w 0-100 16w 0-50 M 16-60 F 11-51 | | <1 day | Contact laboratory. Send to lab IMMEDIATELY Recommended within 15mins |
| Amylase | Serum (Yellow) | 28-100U/L Up to 4 wks old: 12 – 21 U/L Up to 11 Mths old: 12 – 92 U/L Up to 15 Yrs old: 12 – 118U/L >15 Years old: 30 – 118 U/L | | <1 day | |
| Androstenedione | Serum (Yellow) | <9 nmol/L | HALO | <2 weeks | Ranges vary with gender and maturity |
| ACE (angiotensin converting enzyme) | Serum (Yellow) | | HALO | <1 day | |

| | | | | | |
|--|---|--------------------------|------|----------|---|
| Anti-Mullerian hormone (AMH) | Serum (Yellow) | See report | HALO | <1 week | Age- and sex-related reference ranges are on paediatric reports. |
| AMH test is used in the assessment of ovarian reserve in women with endometriosis, or being assessed for assisted reproduction. It may also be used in the assessment of disorders of sexual development in children. | | | | | |
| Results for adult females include interpretation in terms of ovarian reserve status, or risk of ovarian hyperstimulation in assisted reproduction. | | | | | |
| This test may only be requested by or on behalf of a specialist Consultant such as Consultant Obstetrician (Fertility Specialist) or Consultant Paediatrician. | | | | | |
| Aquaporin 4 antibodies | Serum (Yellow) | | HALO | <2 weeks | |
| Aripiprazole | EDTA (Lavender) | | KIT | 2 weeks | By prior arrangement with consultant only |
| This test is occasionally needed for dose adjustment in patients prescribed anti-psychotics, who are on relevant co-medication, smoker etc or who are prescribed off-licence doses | | | | | |
| Arsenic [blood] | Whole blood (EDTA trace element tube (royal blue), EDTA (Lavender) or Heparin (green top) | See report | CHX | 2 weeks | Dietary restrictions required for 5 days prior to sampling. Prior discussion with Biochemistry consultant is advised. |
| Arsenic test is used investigate patients with signs/symptoms of possible arsenic toxicity. The test does not differentiate between toxic inorganic arsenic and non-toxic organic arsenic derived from food. In the UK population, most arsenic is absorbed from food and is not a risk to health. | | | | | |
| Patient must abstain from eating fish, shellfish, seaweed, chicken, rice products, and nutritional supplements for five days before the sample is collected. | | | | | |
| There is PHE guidance at https://www.gov.uk/government/publications/arsenic-properties-incident-management-and-toxicology/arsenic-general-information | | | | | |
| AST (aspartate aminotransferase) | Serum (Yellow) | F 0-32 U/L M 0-40 U/L | | | |
| B2M (beta-2 microglobulin) | Serum (Yellow) | | | | |
| Beta carotene | Serum (Yellow) or Plasma [lithium heparin, Green] | | STH | | Send to lab promptly for immediate freezing of plasma |
| Bicarbonate (HCO3) | Serum (Yellow) | 22-29mmol/L | | <1 day | |
| Bile acids | serum (Yellow) | | | <1 day | |

| Bilirubin (total) | Serum (Yellow) | <21umol/L See Comments section for Age related refs | | <1 day | Age | Lower Limit | Upper Limit |
|------------------------------|---------------------|--|---|----------|--|-------------|-------------|
| | | | | | 0 to < 15 days | 0 | 250 |
| | | | | | 15 days to < 1 year | 0 | 10 |
| | | | | | 1 to < 9 years | 0 | 5 |
| | | | | | 9 to < 12 years | 0 | 8 |
| | | | | | 12 to < 15 years | 0 | 10 |
| | | | | | 15 to < 19 years | 0 | 12 |
| | | | | | ≥ 19 years (Roche) | 0 | 21 |
| Bilirubin (conjugated) | Serum (Yellow) | 0-5umol/L | | <1 day | | | |
| Blood gases (arterial blood) | Heparinised syringe | pH 7.35-7.45 pCO2 4.7-6.0 kPa pO2 10.7-14.6 kPa | | <1 hour | Seal with cap provided. Do not send via pneumatic chute. Label clearly. | | |
| BNP | Serum (Yellow) | See report | | <1 day | | | |
| Bone profile | Serum (Yellow) | | | <1 day | See calcium, phosphatase, albumin & ALP | | |
| C-peptide (of insulin) | Serum (Yellow) | Interpretation with report | HALO / GOS | 1 week | Send to lab promptly for immediate freezing of serum. Assayed only if corresponding glucose <2.5 mmol/l HALO for adult GP samples GOS for paed samples | | |
| CRP (C reactive protein) | Serum (Yellow) | <5mg/L | | <1 day | | | |
| CA125 | Serum (Yellow) | 0-23 IU/L | HALO | <3 days | | | |
| CA153 | Serum (Yellow) | 0-35 IU/l | HALO | <3 days | Not recommended for diagnosis | | |
| CA199 | Serum (Yellow) | 0-35 IU/L | HALO | <3 days | Not recommended for diagnosis | | |
| Caeruloplasmin | Serum (Yellow) | 0.2-0.4g/L | HALO | <2 weeks | | | |
| Calcitonin | Serum (Yellow) | Male <11.8ng/L Female <4.8ng/L | CNIN - (Lister local code/Non-GP) CAC (HSL) - GPs only | <2 weeks | Send to lab promptly for immediate freezing of serum. Fasting sample preferred. | | |
| Adjusted Calcium | Serum (Yellow) | 2.2-2.6 mmol/L Age related ranges in comments | | <1 day | Up to 4 weeks old = 2.0 – 2.7 mmol/L Up to 15 years old = 2.2 – 2.7 mmol/L | | |

| | | | | | |
|---|----------------------------|--|------|-------------|---|
| Carbamazepine | Serum (Yellow) | 4-12ug/L | | <3 days | Collect trough sample just before oral dose |
| Carboxyhaemoglobin | Heparinised syringe | Non smoker 0.5-1.5% Smoker upto 9% | | <1 day | Seal with cap provided. Do not send via pneumatic chute. Label sample. |
| Catecholamines [PLASMA] | Lithium heparin (Green) | This test is not routinely offered. It is not a sensitive test for phaeochromocytoma – send for plasma metadrenalines instead. | | | |
| CEA (carcino embryonic antigen) | Serum (Yellow) | | HALO | <3 days | Higher values in smokers |
| Chloride | Serum (Yellow) | 95-108 mmol/L | | <1 day | |
| Cholinesterase (acetyl) | EDTA (Lavender) | Interpretation with report | HALO | 2 - 3 weeks | For monitoring organophosphate exposure |
| Cholinesterase (pseudo) | EDTA (Lavender) | Interpretation with report | HALO | 2 – 3 weeks | For assessment of succinylcholine sensitivity |
| Clomipramine | EDTA (Lavender) | Interpretation with report | CAR | 1 week | By prior arrangement with consultant only |
| Chromium | Trace metal (Dark Blue) | Interpretation with report | HALO | 1 week | For use of Orthopaedic teams monitoring Metal-on-Metal hip replacements only. Patients consulting GPs with concerns should be referred back to the responsible surgeon. |
| Chromogranin A & B [Part of gut hormone profile] | EDTA (Purple) | Interpretation with report | CHX | 4 weeks | Send to lab promptly for immediate freezing of plasma. |
| Citalopram | EDTA (Lavender) | Interpretation with report | HALO | 1 week | |
| Copper | Trace metal (Dark Blue) | 12-20umol/L | HALO | < 2 weeks | Increased in oral contraceptive use, inflammation Age related ref ranges? 3m (1.4-7.2) 4m(3.9-17.3) 6m11.1-27.4) 9 yrs (11.3-23.7) 13yrs (9.1-22.5) |
| Cortisol | Serum (Yellow) | Interpretation with report | | 1 day | |
| Cortisol test is used for diagnosis and monitoring of adrenal insufficiency. Diagnostic samples are best collected at 8 - 9 am. Due to diurnal rhythm, samples collected at other times have little diagnostic value. | | | | | |

Adrenal insufficiency is excluded in most people if result is above 374nmol/L at any time. For samples collected at 8 – 9 am, a result below 100nmol/L is strongly suggestive of adrenal insufficiency unless there is recent use of corticosteroids, please discuss with Endocrinology.

To confirm adrenal insufficiency, see Short Synacthen test.

This test is not recommended for suspected Cushing's syndrome; please request post-dexamethasone cortisol if performing any dexamethasone suppression test.

This method has improved specificity for cortisol, hence diagnostic action limits are lower than with previous supplier's method. However, there is positive interference from prednisolone, prednisone, methyl prednisolone, and from abnormally raised levels of 11-deoxycortisol or 21-deoxycortisol.

High-dose biotin supplementation (>5mg/day), or increases in cortisol-binding globulin (eg. oral oestrogen therapy, pregnancy) may cause falsely high cortisol results.

| | | | | | |
|--------|-------------------------|--|------|--------|---|
| Cobalt | Trace metal (Dark Blue) | | HALO | 1 week | For use of Orthopaedic teams monitoring Metal-on-Metal hip replacements only. Patients consulting GPs with concerns should be referred back to the responsible surgeon. |
|--------|-------------------------|--|------|--------|---|

| | | | | | |
|---------------------|------------|--|--|----|------------------------------------|
| Covid-19 PCR (POCT) | Viral swab | | | 4h | Follow Trust POCT approval pathway |
|---------------------|------------|--|--|----|------------------------------------|

Covid-19 PCR (POCT) is for the rapid detection of current infection with SARS-CoV-2 virus. Please follow the Trust's current guidance (see Daily Brief) on eligibility and approval for this rapid test or requests will be rejected. The test is available for use with swabs collected from nose, throat, or nasopharynx only.

Please answer the questions appropriately when making ICE requests as this will direct samples either to this low capacity rapid pathway or to the high capacity but slower process provided from Cambridge. Tests required on nasopharyngeal aspirates or other fluids must be processed at Cambridge.

| CK (Creatine Kinase) | Serum (Yellow) | 40-320 (male), 25-200 female U/L | <1 day | Age Range | Sex | CK Reference Range (U/L) |
|----------------------|----------------|--|--------|--------------------|-----|--------------------------|
| | | | | Up to 11 Mnths old | F | 42-493 |
| | | | | Up to 11 Mnths old | M | 39-320 |
| | | | | Up to 15 Years old | F | 39-191 |
| | | | | Up to 15 Years old | M | 43-178 |
| | | | | >15 Yrs old | F | 25-200 |
| | | | | >15 Yrs old | M | 40-320 |

| Creatinine | Serum (Yellow) | M 59-104 µmol/L F 45-84 µmol/L | <1 day | Age | Ref Range(um/L) |
|------------|----------------|-----------------------------------|--------|---------------|-----------------|
| | | | | Up to 2 mnths | 27-87 |
| | | | | Up to 1 yr | 14-34 |
| | | | | 1-2 yrs | 15-31 |
| | | | | 3-4 yrs | 23-37 |
| | | | | 5-6 yrs | 25-42 |

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| | | | | | 7-8 yrs 9-10 yrs 11-12 yrs 13/14 yrs >15 yrs Male >15 yrs Female | 30-47 29-56 39-60 40-68 59-104 45-84 |
| Cystic fibrosis (genetic test) | EDTA (Lavender) | | AGH | 3 - 4 weeks | After positive sweat test. Contact lab | |
| Cycloserine | Serum (Yellow) | | AML | 72hrs from receipt in referral lab | Recommend a pre-dose sample and a post-dose sample, taken 3-4 hours after oral administration. | |
| Cyclosporin | EDTA (Lavender) | 0.55- 1.15ng/ml 51yrs 0.63- 1.44 | HALO | <3 days | Collect trough sample just before oral dose | |
| 11 deoxycortisol | Serum (Yellow) | 7-16 nmol/L | KCH | 2 – 3 weeks | | |
| 7-Dehydro cholesterol | Lithium Heparin (Green) | 7-16mmol/L | Institute of Child health, Guildford | 6-8 weeks | | |
| 5-Alpha Dihydrotestoster one | serum (Yellow) | Interpretation with report | KCH | | Not a first-line test, rarely needed. | |
| Dexamethasone | Serum (Yellow) | Interpretation with report | MWY | 72 hrs | Requestable only by laboratory staff | |

This test is used to assess whether or not adequate dexamethasone blood levels were achieved during an overnight dexamethasone suppression test.

When patients fail to suppress cortisol to <50nmol/L (9am sample, after overnight dexamethasone), the laboratory will arrange for dexamethasone blood level to be measured on that sample.

Dexamethasone ≥3.0 nmol/L is consistent with adequate absorption and metabolism of dexamethasone.

Dexamethasone <3.0 nmol/L suggests impaired absorption or excess metabolism of dexamethasone, an alternative screening test for hypercortisolism is required. It may also indicate dexamethasone has not been taken if non-compliance is suspected.

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|---------------|--------------------|--|------|--------|---|
| DHEA sulphate | Serum (Yellow) | Age related Interpretation with report | CHX | 1 week | Lower levels with increasing age |
| Digoxin | Serum (Yellow) | 0.5-2.0ug/L | | <1 day | Collect trough just before dose. In toxicity sample at 6 hours post dose. Toxicity occurs at lower levels in old age, hypothyroidism, hypokalaemia, hypercalcaemia and hypomagnesaemia. |
| Dothiepin | EDTA (Lavender) | 25-35 | HALO | 1 week | |

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| DPD gene (dihydro-pyrimidine dehydrogenase) | EDTA (blue) | Interpretation with report | AGH | 1 week | Only available to Oncology teams |
| DPD gene test screens for 4 variants of the DPD gene that reduce the enzyme activity and are associated with fluoropyrimidine toxicity in chemotherapy (eg. 5-fluorouracil, capecitabine, tegafur). Results should be used to reduce dose or select alternative chemotherapy regimen as clinically appropriate. Please note that the absence of any of these 4 genetic variants does not eliminate the risk of toxicity. | | | | | |
| The test should only be carried out once, prior to the start of such therapy. Do not repeat the request unless requested to do so by the Laboratory because of a problem with the first sample. | | | | | |
| This test is not available to screen patients prior to use of topical fluorouracil cream as systemic absorption is too low to cause toxicity. | | | | | |
| Ethylene glycol | EDTA (Lavender) | Interpretation with report | SAN | <3 days | Use admission blood sample |
| Ethylene glycol test is used in the diagnosis and monitoring of suspected toxic alcohol poisoning. The test also detects methanol. Other blood tube types may be used if the preferred sample is not available, as long as they do not contain gel. | | | | | |
| The service is available 24/7 for approved diagnostic samples only; please discuss first with Biochemistry Consultant. Please be aware that the presence of ethylene glycol metabolites in blood causes significant elevation in apparent measured lactate, whether measured using a blood gas analyser or the laboratory method. Hence, ethylene glycol poisoning can masquerade as lactic acidosis. | | | | | |
| FK506 (Tacrolimus) | EDTA (Purple) | Interpretation with report | HALO | <3 days | Collect trough sample just before oral dose |
| Ferritin | Serum (Yellow) | (M) 300-400 µg/L (F) 13-150 µg/L (F over 60y) 300-400 µg/L | | <1 day | |
| Ferritin test is used in the investigation of anaemia and the diagnosis and monitoring of iron-deficiency or iron-overload states. | | | | | |
| Ferritin <12 µg/L indicates absent iron stores, however ferritin is a positive acute-phase reactant and may be normal or even raised in the absence of iron stores. | | | | | |
| High ferritin is commonly caused by chronic inflammation, liver disease, kidney disease, alcohol excess, metabolic syndrome, malignancy. These are all more common as causes of high ferritin than is genetic haemochromatosis. If ferritin is above 1200 µg/L, the patient should be investigated for potential iron overload. | | | | | |
| The above reference ranges do not apply to the newborn period, infants or children. | | | | | |
| High-dose biotin supplementation (>5mg/day) may cause falsely low ferritin results. | | | | | |
| FSH (Follicle stimulating hormone) | Clotted (Yellow) | Interpretation with report | | <3 days | Sample on day 1-3 of cycle. State LMP or amenorrhoea. |
| Folate | Serum (Yellow) | >4.5 µg/L | | <1 day | |
| Serum folate test is used in the diagnosis of folate or vitamin B ₁₂ deficiency; a result <3 µg/L is usually taken to indicate deficiency. Serum folate reflects recent folate status and intake, and a folate-rich meal may transiently elevate a low folate into the reference range. If doubt exists, repeat the test in the fasting state. | | | | | |
| High-dose biotin supplementation (>5mg/day) may cause falsely high folate results. | | | | | |
| It is usual to test folate at the same time as vitamin B ₁₂ due to their shared metabolism and the similar clinical features of deficiency (anaemia, macrocytosis, neurological changes). | | | | | |
| Flecainide | EDTA (Lavender) | 0.15-0.9 mg/L | HALO | 1 week | |

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| Fluoxetine | EDTA (Lavender) | 150-500 ug/L | HALO | 1 week | |
| Fluvoxamine | EDTA (Lavender) | 0.1-0.5 mg/L | HALO | 1 week | |
| Free Fatty Acids | Lithium Heparin (Green) | Interpretation with report | GOS | <2 weeks | Send to lab promptly for immediate freezing of plasma |
| Fructosamine | Serum (Yellow) | 0-282 μ mol/L | HALO | 1 week | Available only with consultant approval |
| fPSA (free PSA) | Serum (Yellow) | <25% | | <1 day | Results expressed as %fPSA/PSA |

This test is used to assist in the detection of prostate cancer.

In men with borderline PSA results and normal DRE, the specificity for detecting prostate cancer may be increased with concurrent use of fPSA. This is expected to detect 92.5% of prostate cancers, and potentially avoid biopsy in 20% of men over 50 years old who do not have prostate cancer.

The method used is Roche COBAS. Free PSA values may vary depending on the method used, and must only be interpreted with total PSA results also produced using Roche COBAS.

High-dose biotin supplementation (>5mg/day) may cause falsely low fPSA results. Always interpret results in the light of the clinical picture.

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| FT3 (free tri-iodothyronine) | Serum (Yellow) | 3.1-6.8 pmol/L | | <1 day | |
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FT3 test is used in the diagnosis/monitoring of subclinical hyperthyroidism. It can also be helpful in monitoring treatment with liothyronine, and the differential diagnosis of thyroiditis/Graves disease. There is no value in measuring FT3 in patients taking thyroxine. FT3 is only requestable for patients under the care of Endocrinology, or may be added at the discretion of Laboratory Consultant.

High-dose biotin supplementation (>5mg/day) may cause falsely high FT3 results. Due to biotin interference, TFTs in euthyroid patients taking very high-dose biotin may falsely resemble the pattern seen in thyrotoxicosis.

Age related reference ranges

| Age | Range (pmol/L) |
|---------------------|----------------|
| 0-<1 month | 4.2-13 |
| 1 month -<12 months | 5.2-8.6 |
| 1 year - <17 years | 5-8.2 |
| >17 years | 3.1-6.8 |

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| FT4 (free thyroxine) | Serum (Yellow) | 12-22 pmol/L | | <1 day | |
|----------------------|----------------|--------------|--|--------|--|

FT4 test is used in the diagnosis and monitoring of thyroid disease, and is requested as part of the TFT profile. Interpret results in the light of the TSH result, and the clinical picture. The above reference range does not apply to the newborn period, infants or in pregnancy.

Many drugs affect FT4 results independent of any real effect on thyroid status:

Low FT4 is seen with carbamazepine, frusemide, salicylate and others.

High FT4 is seen with amiodarone, heparin.

High-dose biotin supplementation (>5mg/day) may cause falsely high FT4 results.

Due to biotin interference, TFTs in euthyroid patients taking very high-dose biotin may falsely resemble the pattern seen in thyrotoxicosis.

After starting thyroxine or changing dose, wait 2-3 months before checking TFTs.

Avoid testing TFTs during moderate-severe illness unless there is strong clinical indication, as transient changes in TFTs are common at this time (euthyroid sick syndrome).

Age related reference ranges

| | | | | | |
|---|-------------------------|--|------|---|---|
| 0 to <1 month | 16 | 50 | | | |
| 1 to <12 months | 14 | 22 | | | |
| 1 to <19 years | 13 | 21 | | | |
| ≥19 (Roche range) | 12 | 22 | | | |
| <hr/> | | | | | |
| Gamma GT | Serum (Yellow) | Male 15-73U/L Female 12-43 U/L | | <3 days | Ranges vary for children. No longer part of LFT. Must be specifically requested. Age related ref ranges Up to 4 Weeks old: 23 - 219 Up to 11 Months old: 8 - 127 Up to 15 Years old: 6 - 21 U Females >15 years old: 0 - 37 U Males >15 Years old: 0 - 72 U |
| Gabapentin | EDTA (Lavender) | 2-20 mg/L | HALO | 7 days | Collect trough sample just before oral dose |
| Galactose-1-Phosphate Uridyl Transferase | Lithium Heparin (Green) | 18-40 µmol/h/gHb | HALO | 4 weeks | |
| Gastrin [Part of gut hormone profile] | EDTA (Lavender) | <40pmol/L | CHX | 4 weeks | Fasting sample required. Send to lab promptly for immediate freezing of plasma |
| Guanidino Acetic Acid [GAA] | Lithium Heparin (Green) | Interpretation with report | ADI | | |
| Gentamicin | Serum without gel (Red) | For once a day dosing Pre dose <1.0mg/L (adults) <2.0mg/L (neonates) | | 2hours from receipt, inform lab if urgent | Inform lab if urgent Refer to antibiotic guidelines on KC for further information For b d dosing eg endocarditis discuss with Cons.Microbiologist |
| Glucose | Fluoride EDTA (Grey) | 3.5-6.0 mmol/L fasting | | <1 working day | |
| Growth Hormone (GH) | Serum (Yellow) | Interpretation with report | CHX | 1 week | Random samples rarely helpful. Use dynamic function tests instead |
| Gut hormone profile (Gastrin, Glucagon, pancreatic polypeptide, somatostatin, vasoactive intestinal peptide & chromogranin A & B) | EDTA (Lavender) | Interpretation on report | CHX | 4 weeks | Fasting sample required. Send to lab promptly for immediate freezing of plasma |

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| HbA1c (glycated Hb) | EDTA (Lavender) | | | <3 days | |
| hCG (human chorionic gonadotrophin) | Serum (Yellow) | 0-4U/l | | <1 day | For diagnosis and monitoring of ectopic pregnancy or germ cell tumour |
| HDL cholesterol | Serum (Yellow) | See guidelines in BNF | | <1 day | |
| HLA B27 | EDTA-(Blue) | | HALO | 14 days | The presence or absence of HLA-B27 is used in the differential diagnosis of a number of inflammatory diseases |
| HMGCR autoantibodies | Serum (Yellow) | Negative = <14.9 CU/ml Equivocal = 15-24.9 CU/ml Positive = >25.0 CU/ml | OXF | 14 days | |
| 17-hydroxy-progesterone (17OHP) | Serum (Yellow) | 0-5nmol/L (adults) | HALO | 2 weeks | Reference ranges vary in children. |

17OHP test is a first-line investigation in the diagnosis of congenital adrenal hyperplasia (CAH) due to deficiency of an enzyme in the steroid synthetic pathway (most commonly 21-hydroxylase, but also 11-hydroxylase and 3-beta hydroxysteroid dehydrogenase deficiency). 21-hydroxylase deficiency is excluded if 17OHP is within the reference range.

17OHP is high at birth and decreases to the reference range after about 48h; to avoid false positives blood should not be collected from newborns before they are 48h old. Classical salt-losing 21-OHylase deficiency usually presents clinically in the second week of life. 17OHP may remain high for longer than 48h in premature babies and term babies who are sick.

In adult women suspected of non-classical CAH, collect blood in the early morning and in the follicular phase (results are higher in the luteal phase).

For full diagnosis, a short Synacthen test is required that measures steroid precursors as well as cortisol.

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| Plasma Homocysteine | Lithium Heparin (Green) | 5-15 µmol/L | QUE | 1-3 weeks | Send to lab promptly for immediate freezing of plasma |
|---------------------|-------------------------|-------------|-----|-----------|---|

Plasma homocysteine test may be used when there is strong clinical suspicion of vitamin B12 deficiency (peripheral axonal neuropathy, subacute degeneration of spinal cord) but serum B12 results are indeterminate (200 - 300ng/L). Homocysteine is typically raised in vitamin B12 deficiency, but also in folate deficiency, vitamin B6 (pyridoxine) deficiency, and several other conditions. It is also increased in renal impairment.

This test may also be used in the diagnosis of homocystinuria.

Do not use this test as a marker of cardiovascular risk.

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|---------------------------------|--------------------------------------|------------------------------|-----|-----------|--------------------------|
| Total β Hexosaminidase | Lithium heparin (Green) | Interpretation with report | GOS | 6-8 weeks | |
| Hexosaminidase A | Lithium heparin, whole blood (Green) | Interpretation with report | GOS | 6-8 weeks | |
| Immunoglobulins (IgG, IgA, IgM) | Serum (Yellow) | Refer to immunology handbook | ADI | <3 days | Ranges vary for children |

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| Insulin | Serum (Yellow) | Interpretation with report | HALO / GOS | 1 week | Send to lab promptly for immediate freezing of serum. Test will only be undertaken if glucose level is <2.5 mmol/l HALO is for adult samples GOS for paed samples |
| Insulin like growth factor (IGF-1) | Serum (Yellow) | 13-64nmol/L | HALO | 1 week | Higher values in children. Age related ranges on reports |
| IGF-Binding Protein 3 | Serum (Yellow) | Interpretation with report | HALO | 1 week (as required) | |
| Inhibin B | Serum (Yellow) | Interpretation with report | HALO | 2 weeks | |
| Imipramine | EDTA (Lavender) | 75-160 μ mol/L | HALO | 1 week | |
| Iron (Fe) | Serum (Yellow) | 5.83-34.5 μ mo/L | | <1 day | Ferritin is better test for iron deficiency |
| Lactate | Fluoride EDTA (Grey) | 0.6-2.5 mmol/l | | <1 day | |
| Please be aware that the presence of ethylene glycol metabolites in blood causes significant elevation in apparent measured lactate, whether measured using a blood gas analyser or the laboratory method. Hence, ethylene glycol poisoning can masquerade as lactic acidosis | | | | | |
| Lactate dehydrogenase (LDH) | Serum (Yellow) | 135-225 IU/L | | <1 day | Ranges vary in children |
| Lamotrigine | EDTA (Lavender) | 3-15 mg/L | HALO | 2-3- weeks | |
| Laxative screen [urine] | Random urine | Interpretation with report | HALO | 1 week | |
| LDL cholesterol | Serum (Yellow) | 0-3 mmol/L | | <1 days | Calculated from other lipid values if triglycerides <4.5 mmol/l |
| Lead (Pb) | EDTA (Lavender) | Environmental exposure <0.6 μ mol/L | HALO | <2 weeks | For guidance on industrial exposure contact laboratory. Test done on whole blood. |
| Levetiracetam | EDTA (Lavender) | 10-37 mg/L | HALO | 2 - 3 weeks | By prior arrangement with consultant only |
| Liver function tests (LFTs) | LFTs comprise ALT, ALP, albumin, bilirubin (total). See individual tests for further information. Repeat testing of LFTs is rarely useful within 72 hours of a previous test, but more frequent monitoring <u>may</u> be needed in acute liver injury, acute paracetamol poisoning, and ICU patients. Please request tests individually if you do not require all 4 results (eg. ALT is usually sufficient when monitoring statin treatment). | | | | |
| Lipid profile (fasting) | Serum (Yellow) | | | <1 day | Fasting sample required (12 h) see cholesterol, HDL |

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| | | | | | cholesterol, LDL cholesterol & triglycerides. |
| Lipoprotein A | Serum (Yellow) | 0-32 nmol/L | HALO | 1 -2 weeks | By prior arrangement with consultant only |
| Free Light Chains | Serum (Yellow) | Interpretation with report | HALO | | For Haematologists and Renal only |
| Lithium | Serum (Yellow) | 0.4-1.0mmol/L | | <1 day | |
| Luteinising hormone (LH) | Serum (Yellow) | Interpretation with report | HALO | <1 day | |
| Magnesium | Serum (Yellow) | 0.7-1.0mmol/L | | <1 day | Age related ref ranges Up to 4 weeks old 0.6-1.0mmol/L >4 weeks old 0.7-1.0mmol/L |
| Manganese | Trace metal (dark blue) | 9-25 nmol/L | HALO | <2 weeks | |
| Mannose Binding Lectin | Serum (Yellow) | 0.7-6.0 mg/L | SHP | 7 working days | |
| This test should only be used if Mannose Binding Lectin (MBL) deficiency is suspected, as it is associated with an increased risk of infection when the adaptive immune system is immature (early childhood), or when it is suppressed (e.g., transplantation, chemotherapy treatment). | | | | | |
| Mercury | EDTA (Lavender) | <30nmol/L | HALO | <2 weeks | Done on whole blood for organic Hg exposure. Urine preferred for inorganic mercury |
| This test should be used only for cases of suspected poisoning with organomercury compounds such as methyl mercury (antifungal compound previously used in agriculture) | | | | | |
| Do not use in relation to dental amalgam fillings. There is no convincing evidence that these cause any adverse health effects. | | | | | |
| Metabolic stone screen (blood) | Serum (Yellow) | | | <1 day | Na+, K+, HCO3, Urea, Creatinine, Albumin, Ca++, PO4 and urate |
| Methaemoglobin | Heparinised syringe | <1.5% | | <1 day | Seal with cap provided. Do not send via pneumatic chute. Label sample. |
| Methylmalonic acid (MMA) | Plasma | In the context of a raised total homocysteine and the absence of renal impairment: < 0.29 mmol/l are considered | QUE | <2 weeks | |

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| | | not indicative of B12 deficiency 0.29-0.70 mmol/l suggests B12 deficiency > 0.70 mmol/l consistent with overt B12 deficiency Supported by in house data | | | |
| <p>Plasma methylmalonic acid test may be used when there is strong clinical suspicion of vitamin B12 deficiency (peripheral axonal neuropathy, subacute degeneration of spinal cord) but serum B12 results are indeterminate (200 - 300ng/L). MMA is typically raised in vitamin B12 deficiency, but not in folate deficiency. It is also increased in renal impairment.</p> <p>It should not be used for diagnosing methylmalonic aciduria as it lacks sensitivity for this purpose, and urinary MMA should be used instead.</p> | | | | | |
| Metadrenalin Blood | EDTA (Lavender) | Interpretation with report | MAN | 2-3 weeks | Send to lab promptly for immediate freezing of plasma. Patient should have indwelling venous catheter and be supine for 30mins before sampling. |
| Myelin Oligodendrocyte (MOG) | Serum (Yellow) | Interpretation with report | ION | <2 weeks | |
| Neurotensin | EDTA (Lavender) | Interpretation with report | CHX | 3 weeks | Send to lab promptly for immediate freezing of plasma |
| Neurone specific enolase (NSE) | Serum (Yellow) | <13 μ g/L | CHX | 4 weeks | |
| Oestradiol | Serum (Yellow) | M28-156 pmol/L | | 1 day | Reference ranges for females are on the report |
| <p>Oestradiol test is for use in investigation of disorders of sexual development, patients using oestradiol implants, gynaecomastia in males, or possible oestrogen secreting tumour. It is not recommended when querying menopausal status.</p> <p>High-dose biotin supplementation (>5mg/day) may cause falsely high oestradiol results. Always interpret results in the light of the clinical picture.</p> | | | | | |
| Olanzapine | EDTA (Lavender) | 20-40 μ g/L | HALO | 2 weeks | By prior arrangement with consultant only |
| <p>This test is occasionally needed for dose adjustment in patients prescribed anti-psychotics, who are on relevant co-medication, smoker etc or who are prescribed off-licence doses.</p> | | | | | |
| Osmolality | Serum (Yellow) | 275-295 mmol/Kg | | <1 day | |
| <p>This test can be useful in the differential diagnosis of hypo- or hyper-natraemia, especially if compared to a urine osmolality collected at the same time. It can also be used in combination with serum Na, K, urea, and glucose results to calculate the osmolal gap, which can aid in the diagnosis of poisoning by toxic alcohol such as methanol, ethylene glycol etc.</p> | | | | | |

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| Oxalate, Blood | EDTA (Lavender) | Interpretation with report | UCLH | | |
| Oxcarbazepine | EDTA (Lavender) | Interpretation with report | HALO | 7 days | By prior arrangement with consultant. |
| Pancreatic polypeptide [Part of gut hormone profile] | EDTA (Lavender) | Interpretation with report | CHX | 4 weeks | Fasting sample required. Send to lab promptly for immediate freezing of plasma. |
| Paracetamol | Serum (Yellow) | 10-30mg/l therapeutic | | <1 day | Overdose – collect at least 4 hours after overdose. See BNF for interpretation and treatment guidelines |
| Parathyroid hormone (PTHi) | EDTA (Lavender) | 1.6-6.9 pmol/L | | <8 days | |
| pH | Pleural Fluid/ Urine | Stated on report | | 30 mins | Must arrive in the laboratory promptly for immediate analysis |
| Phenobarbitone | Serum (Yellow) | 10-40mg/L | | <3 days | |
| Phenylalanine | Blood spot card or Lithium heparin can be used | 55-206µmol/L | GOS | 1-2 weeks | Send to lab promptly for immediate freezing of plasma |
| Phenytoin | Serum (Yellow) | 5-20mg/L | | <3 days | |
| Phosphate (PO4) | Serum (Yellow) | 0.8-1.5mmol/L | | 1 day | Ranges vary in children |
| Placental growth factor (sFlt1/PIGF ratio) | Serum (Yellow) | 0-37ng/L | | <1 day | |
| This test is offered for the rapid rule-out of pre-eclampsia for up to 1 week, in pregnant women between 20 and 35 weeks gestation. | | | | | |
| Post-dexamethasone cortisol | Serum (Yellow) | <50nmol/L | | 1 day | |
| This test is used to exclude hypercortisolism (Cushing's syndrome) in patients at low risk. Dexamethasone should be taken at 11pm - 12 midnight. Collect blood sample at 8 - 9 am next morning. See cortisol test for important information about limitations of the cortisol assay. | | | | | |
| Potassium | Serum (Yellow) | 3.5-5.3mmol/L | | <1 day | Values increased by haemolysis and prolonged contact with red cells |
| Procalcitonin | Serum (Yellow) | 0-0.05µg/L | | <4h | |
| Procalcitonin test is used to aid in deciding whether an infection is viral or bacterial, and can also be used to guide the need for antibiotic therapy. Currently only available for requesting by ICU Consultants, or others after approval by Consultant Microbiologist. Result <0.5 µg/L represents a low risk of severe sepsis and/or septic shock | | | | | |

| Result >2.0 µg/L represents a high risk of severe sepsis and/or septic shock | | | | | |
|--|--|---|------|-------------|---|
| Porphyrins | EDTA (Lavender) | Interpretation with report | BED | 1 week | For detection of porphyrias with non-acute presentation. Exclude light from samples. |
| Procollagen III peptide (P3NP, PIII NP) | Serum (Yellow) | 1.2-4.2µg/L | HALO | 2-3 weeks | Requested by dermatology only |
| Procainamide | EDTA (Lavender) | Interpretation with report | HALO | 7 Days | |
| Progesterone | Serum (Yellow) | Day 21 3-95nmol/L | | <3 days | Record day of cycle and treatment. Ovulation likely if mid luteal phase level >30 mmol/l |
| Prolactin | Serum (Yellow) | Male 86-324 mIU/L Female 102-496 mIU/L | | <3 days | |
| Protein electrophoresis | Serum (Yellow) | Interpretation with report | HALO | <7 days | Early morning urine for BJP is required as part of the myeloma screen |
| Prostate Specific Antigen (PSA) | Serum (Yellow) | Age-related ranges, see comments | | <1 day | <40y 0-1.4 µg/L 40-49y 0-2.0 µg/L 50-59y 0-3.1 µg/L 60-69y 0-4.1 µg/L ≥70y 0-4.4 µg/L |
| <p>This test is used in the detection and monitoring of prostate cancer. In men with borderline results and normal DRE, the specificity for detecting prostate cancer may be increased with concurrent use of fPSA.</p> <p>The method used is Roche COBAS. PSA values may vary depending on the method used.</p> <p>High-dose biotin supplementation (>5mg/day) may cause falsely low PSA results. Always interpret results in the light of the clinical picture.</p> | | | | | |
| Pyruvate | Special tube needed. Discuss with lab | <0.18 mmol/L | GOS | 1 - 2 weeks | By prior arrangement with consultant |
| Quetiapine | EDTA (Lavender) | | KIT | 2 weeks | By prior arrangement with consultant only |
| <p>This test is occasionally needed for dose adjustment in patients prescribed anti-psychotics, who are on relevant co-medication, smoker etc or who are prescribed off-licence doses.</p> | | | | | |
| Renal function tests | Renal function tests comprise Na, K, urea, creatinine, eGFR. See individual tests for further information. | | | | |
| Renin | EDTA (Lavender) | | ADI | 2 weeks | Please note this is a mass assay and not an activity assay. |

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| Retinol Binding Protein (Urine) | Fresh random urine | Interpretation with report | GOS | 1-3 weeks | Freeze soon after collection and send frozen. |
| Risperidone | EDTA (Lavender) | | KIT | 2 weeks | By prior arrangement with consultant only |
| This test is occasionally needed for dose adjustment in patients prescribed anti-psychotics, who are on relevant co-medication, smoker etc or who are prescribed off-licence doses. | | | | | |
| Salicylate | Serum (Yellow) | >300mg/l toxic | | <1 day | See BNF for guidelines |
| Selenium | Trace metal (dark Blue) | 0.85-1.46 µmol/L | HALO | <2 weeks | |
| Sertraline | EDTA plasma (trough level) | With report | HALO | 7 days | |
| Sex hormone binding globulin (SHBG) | Serum (Yellow) | | HALO | 1 week | |
| Sirolimus (Rapamune) | EDTA (Lavender) | Interpretation with report | HALO | <3 days | Collect trough sample just before oral dose |
| Sodium (Na+) | Serum (Yellow) | 133-146 mmol/L | | <1 day | |
| Somatostatin [part of gut hormone profile] | EDTA (Lavender) | Interpretation with report | CHX | 2 -3 weeks | Fasting sample required. Send to lab promptly for immediate freezing of plasma |
| Squamous cell carcinoma antigen (SCC) | Serum (Yellow) | <150ng/dL | HALO | <1 week | |
| Steroid Profile | Random Urine. | Interpretation with report | UCLH | 2 -3 weeks | |
| Sulpiride | EDTA (Lavender) | | KIT | 2 weeks | By prior arrangement with consultant only |
| This test is occasionally needed for dose adjustment in patients prescribed anti-psychotics, who are on relevant co-medication, smoker etc or who are prescribed off-licence doses. | | | | | |
| Sulphonylurea | Serum (Yellow) | Interpretation with report | RSC | 2 weeks | |
| Short Synacthen test | Serum (Yellow) | Interpretation with report | | 1 day | Samples to be collected at 0 mins, 30 mins and 60 mins post-synacthen injection. |
| Short Synacthen test (SST) is used in diagnosis of adrenal insufficiency. In patients already taking corticosteroids, postpone SST until at least 18-24h after the last dose, and longer if taking synthetic corticosteroids. It is usually helpful to collect blood for ACTH test before the Synacthen injection. Stimulated cortisol result should be above 374nmol/L See cortisol test for important information about limitations of the cortisol assay. | | | | | |
| Tacrolimus (FK506) | EDTA (Lavender) | Interpretation with report | HALO | <3 days | Collect trough just before oral dose |

| | | | | | |
|---|-----------------------------|--|------|----------|---|
| | | | | | |
| Teicoplanin | Serum (Yellow) | Interpretation with report | HALO | <5 days | Collect trough sample just before next dose |
| Testosterone | Serum (Yellow) | 0.1-1.7 nmol/L (adult female). Age-related ranges for adult males, see comments | | <3 days | Adult male reference ranges 19 to 39y 8.0 – 31.3 40 to 49y 7.2 – 31.3 50 to 59y 6.7 – 31.3 60 to 79y 6.6 – 31.3 80 to 99y 4.1 – 31.3 |
| <p>This test is used in the diagnosis and monitoring of disorders of androgen production, disorders of sexual development, and for monitoring androgen suppression therapy in prostate cancer.</p> <p>In adult men there is a wide diurnal variation in testosterone levels; the quoted reference ranges assume phlebotomy at around 09:00am (\pm 2h). Results from samples collected in the late afternoon from healthy young men may be only about half of those found in 09:00am samples, and are not easily interpreted.</p> <p>For children, reference ranges based on Tanner stage are printed on the report.</p> <p>High-dose biotin supplementation (>5mg/day) may cause falsely high testosterone results. Always interpret results in the light of the clinical picture.</p> | | | | | |
| TFTs (thyroid function tests) | Serum (Yellow) | TFT profile comprises TSH and FT4. See individual tests for more information. | | | |
| Theophylline | Serum (Yellow) | 10-20mg/L | | <1 day | Oral sustained release – collect before dose. IV – collect 6 and 18hrs post dose. |
| Thiopentone | EDTA (Lavender) | Interpretation with report | MTL | <24 hrs | |
| Thiopurine metabolites (6-TGNs) | EDTA whole blood (red) | 235-450 pmol/8x10 ⁸ RBC | SAN | < 1 week | |
| <p>This test is used in patients with a poor response to thiopurine drugs, to distinguish those with inadequate dosing or poor adherence to medication from those who are resistant to the drug and need alternative therapy (~20% of those with normal TPMT).</p> <p>Samples should be collected no less than 4 weeks after initiation of therapy, or a change in dose. The t½ is several days so samples can be collected at any time in relation to the dose.</p> <p>6-methylmercaptopurine nucleotides (6-MMPNs) are also reported (reference range <5700 pmol/8x10⁸ RBC). 6MMPNs are inactive in terms of therapeutic effect, but are hepatotoxic.</p> <p>Although a mixture of molecules, 6-TGNs correlate well with clinical response and can be used to some extent in dose optimisation.</p> | | | | | |
| TPMT (thiopurine methyl transferase) | EDTA whole blood (Lavender) | 68-150 mU/L | SAN | < 1 week | |
| <p>TPMT test is used to guide dosage of thiopurine drugs such as azathioprine. Guidelines recommend assessing TPMT status before starting treatment.</p> <p>These drugs are metabolised to inactive products by TPMT. 11% of the population has low levels of TPMT so is more susceptible than normal to toxic effects such as myelosuppression. Note that TPMT status fails to predict the majority of myelosuppression with these drugs, and ongoing FBC and LFT monitoring remains pivotal to safe therapy.</p> <p>Blood transfusion within the last 90 days may misclassify TPMT status and mask a deficient TPMT result.</p> <p>This test assesses the patient's phenotype, hence it should not generally ever be repeated.</p> | | | | | |
| Thyroglobulin | Serum (Yellow) | After total thyroidectomy or I ¹³¹ | HALO | ~8 days | Includes thyroglobulin antibodies (TgAb) |

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| | | ablation: <0.1 μ g/L Birth-3 weeks: 10-250 μ g/L | | | |
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Thyroglobulin test is used to assess whether any thyroid tissue remains after total thyroidectomy or I^{131} ablation, in patients with known thyroid cancer. It may rarely be used to assess newborns with suspected thyroid development problems.

Because endogenous thyroglobulin antibodies (TgAb) can interfere with thyroglobulin assays, TgAb are also reported at the same time.

High-dose biotin supplementation (>5mg/day) may cause false thyroglobulin results. Always interpret results in the light of the clinical picture.

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|------------------------|-------------------------------------|----------------------------|------|-------------------|---|
| Topiramate | EDTA (Lavender) | Interpretation with report | HALO | 7 days | By prior arrangement with consultant only |
| Total protein (TP) | Serum (Yellow) | 60-80g/L | | <1 day | |
| Transferrin | Serum (Yellow) | 2-3.4 g/L | | <1 day Mon- Fri | |
| Transferrin Glycoforms | Serum (Yellow)/heparin Plasma 0.5ml | Interpretation with report | ION | <10 days | |
| Triglycerides | Serum (Yellow) | 0.3-1.8mmol/L | | <1 day | See also lipids – fasting specimen required |
| Troponin T | Serum (Yellow) | 0-14 ng/L | | <4h <1h for ED | |

This test is used to assess the risk of acute coronary syndrome or myocardial infarction in patients with ischaemic symptoms.

High-dose biotin supplementation (>5mg/day) may cause falsely low troponin T results. Always interpret results in the light of the clinical picture.

The above reference range does not apply to dialysis patients, newborns, or infants.

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| Tryptase | Serum (Yellow) | 2-14 μ g/L | HALO | <1 week | |
|----------|----------------|----------------|------|---------|--|

Tryptase test is used mainly in the investigation of suspected anaphylaxis, but may rarely be required in the investigation of systemic mastocytosis. Tryptase, histamine, and other mediators of allergic response are released from mast cell granules when mast cells are activated.

For suspected anaphylaxis, serial samples should be collected to follow the kinetics of release: within 1h of reaction, and subsequently at 3h and 24h. Peak levels of >40 μ g/L are associated with anaphylaxis.

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|-----------------------------------|----------------|---------------|--|--------|--|
| TSH (thyroid stimulating hormone) | Serum (Yellow) | 0.27-4.2 mU/L | | <1 day | |
|-----------------------------------|----------------|---------------|--|--------|--|

TSH test is used in the diagnosis and monitoring of thyroid disease. It is usually requested as part of the TFT profile. Interpret results in the light of the FT4 result, and the clinical picture. The above reference range does not apply to the newborn period, infants or in pregnancy.

In the special case of monitoring stable patients taking thyroxine, TSH alone may be requested without FT4.

High-dose biotin supplementation (>5mg/day) may cause falsely low TSH results. Due to biotin interference, TFTs in euthyroid patients taking very high-dose biotin may falsely resemble the pattern seen in thyrotoxicosis.

After starting thyroxine or changing dose, wait 2-3 months before checking TFTs.

Avoid testing TFTs during moderate-severe illness unless there is strong clinical indication, as transient changes in TFTs are common at this time (euthyroid sick syndrome).

Age related reference ranges are reported.

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|--|----------------|--|--|----------|---|
| Urea & electrolytes (U/E) | Serum (Yellow) | | | <1 day | See NA+, K+, Urea & Creatinine |
| Urate | Serum (Yellow) | 200-430 male 140-360 female μ mol/L | | <1 day | |
| Urea | Serum (Yellow) | 2.5-7.8mmol/L | | <1 day | |
| Valproate, Valproic acid, Sodium Valproate | Serum (Yellow) | | | <3 days | Only available for neurology and psychiatry unless discussed with laboratory consultant Valproate is not a useful marker of treatment efficacy: use only to confirm compliance or self poisoning |
| Vancomycin | Serum (Yellow) | 10-20 mg/L | | <8 hours | |

This test is used to optimise dosage to achieve maximal antimicrobial effect (ensure levels are at least 10mg/L) whilst minimising the risk of nephrotoxicity.

For severe infections involving MRSA pneumonia, osteomyelitis, endocarditis, and bacteraemia, adjust dose to achieve an optimal target concentration of 15-20mg/L.

Further interpretation is given on the report.

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|---|--------------------------------|----------------------------|------|-------------|--|
| Venlafaxine | Serum [no gel separator tubes] | Interpretation with report | HALO | 1 week | Only by prior arrangement with consultant |
| Vasoactive intestinal polypeptide (VIP) [part of gut hormone profile] | EDTA (Lavender) | <30pmol/L | CHX | 4 weeks | Send to lab promptly for immediate freezing of plasma. |
| Very long chain fatty acids (VLCFA) | Lithium Heparin (Green) | Interpretation with report | GOS | 2 - 4 weeks | Send to lab promptly for immediate freezing of plasma A fasting sample is preferred |

This test is used in the investigation of peroxisomal disorders where there is a defect in the metabolism or processing of very long chain fatty acids (i.e. fatty acids with a carbon length >22). The test is particularly useful for the diagnosis of X-linked adrenoleukodystrophy in males. However, a normal very long chain fatty acid profile may be seen in asymptomatic female carriers. The test also includes pristanate and phytanate measurement which are useful for the diagnosis of Refsum disease, methyl-acyl CoA racemase deficiency and rhizomelic chondrodyplasia punctata (depending on the age of the patient). A fasted sample is preferred due to dietary influence on VLCFA levels.

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| Vigabatrin | EDTA (Lavender) | 5-35mg/L therapeutic | HALO | 2-3 weeks | |
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|---|---|---------------------------------------|------|-------------|---|
| Vitamin A | Serum [Yellow] or Lithium Heparin (Green) | 1.1-2.8 μ mol/L | GOS | 2-4 weeks | Protect sample from light. Send to lab promptly for immediate freezing of plasma |
| Vitamin B ₁₂ | Serum (Yellow) | 200-770 ng/L | | <1 day | |
| <p>Vitamin B₁₂ test is used in the diagnosis of vitamin B₁₂ or folate deficiency. In the presence of strong clinical suspicion, low serum vitamin B₁₂ (<200 ng/L) is consistent with deficiency. Normal vitamin B₁₂ does not exclude deficiency if there are strong clinical features; to avoid neurological impairment, treatment should not be delayed.</p> <p>Vitamin B₁₂ falls by 30% by the third trimester of pregnancy. In the absence of strong clinical suspicion, slightly low vitamin B₁₂ (150-200ng/L) may be physiological in late pregnancy.</p> <p>Repeat testing is not needed in patients with vitamin B₁₂ deficiency.</p> <p>In subclinical deficiency (vitamin B₁₂ level 150-200ng/L in the absence of symptoms), vitamin B₁₂ level may be checked after 2 months.</p> <p>High-dose biotin supplementation (>5mg/day) may cause falsely high vitamin B₁₂ results.</p> <p>It is usual to test folate at the same time as vitamin B₁₂ due to their shared metabolism and the similar clinical features of deficiency (anaemia, macrocytosis, neurological changes).</p> | | | | | |
| 1,25 dihydroxyvitamin D. Also a pop up in ICE. | Serum (Yellow) | Adult (> 18 yr) 55 – 139 pmol/L | HALO | 4 weeks | Children ref ranges: 0 – < 1 yr 77 – 471 pmol/L 1 – < 3 yr 113 – 363 pmol/L 3 – < 19 yr 108 – 246 pmol/L |
| <p>1,25 dihydroxyvitamin D.</p> <p>This test is very rarely indicated, such as cases of suspected hereditary rickets. Request it only after discussion with Biochemistry clinical team.</p> <p>To assess vitamin D status, please request vitamin D (25-hydroxyvitamin D) instead.</p> | | | | | |
| Vitamin D (25OHD) | Serum (Yellow) | > 50nmol/L | HALO | 1 - 2 weeks | 25-50nmol/L may be inadequate in some people. < 25nmol/L: treatment recommended |
| <p>Vitamin D test is used in the diagnosis of vitamin D deficiency or, rarely, toxicity. Asymptomatic individuals at increased risk of vitamin D deficiency (elderly, dark skin etc) should take a daily supplement of 400 IU, they do not need a vitamin D test.</p> <p>Vitamin D test is recommended in patients with bone diseases that may improve with vitamin D treatment (osteomalacia, osteoporosis, hyperparathyroidism). It is also appropriate in those with chronic musculoskeletal pain or other symptoms possibly attributable to osteomalacia.</p> <p>Patients taking vitamin D supplements do not need routine vitamin D monitoring, except perhaps those with symptomatic deficiency, malabsorption, or where poor adherence is suspected. Any repeat test should be at least 3 months after dosage change.</p> <p>Do not measure vitamin D in patients taking alfalcacidol or calcitriol; these substances are not measured by the assay.</p> <p>It is recommended to check serum calcium one month after starting vitamin D supplements, in case previously undiagnosed hyperparathyroidism is unmasked.</p> | | | | | |
| Vitamin E (tocopherol) | Serum (Yellow) | 11.5-35.0 μ mol/L | ION | 2 - 4 weeks | |
| Zinc | Trace metal (orange) | 9.8-19 μ mol/L | HALO | <2 weeks | |

Biochemistry urine tests

If the assay required is **NOT** listed then please contact the department directly. All results are reported with reference ranges and/or an interpretative comment. Advice on appropriate requesting and interpretation of results is available at all times from the Chemical Pathologists and Clinical Biochemists.

Indicative turnaround times are given for non-urgent situations. It may be possible to expedite results by discussing the clinical situation with the laboratory consultant staff.

If a required test is not listed here, please discuss with the laboratory consultant staff.

For full referral address refer to Appendix 1 (Blood sciences)

| Test | Sample | Adult Reference range | *If referred site code | Turnaround time | Comments |
|---|--|-------------------------------------|---------------------------|--------------------|--|
| 5-hydroxyindole acetic acid (5-HIAA) | 24hr collection into 25 mL 5M HCL | 0-50 µmol/24hr | MWY | 2-3 weeks | For 48h before and during the collection, avoid eating banana, tomato, walnut, aubergine, avocado, pineapple |
| Urine 5-HIAA test is now replaced by serum 5-HIAA, which is simpler and more convenient for patients, does not require 3 days of dietary restrictions, and avoids risk of harm to patients associated with the strong acid in the collection bottle. Please use the serum test instead or discuss with Biochemistry Consultant if you feel the urine test is still warranted. | | | | | |
| Albumin (microalbumin) | Random urine - plain | Reported as ACR, see comments | | <1 day | Reference ranges: <3.5mg/mmol creatinine (female) <2.5mg/mmol creatinine (male) |
| Urine albumin test is an early and sensitive marker for chronic kidney disease. It can be temporarily raised by UTI, physical illness, exercise, diurnal variation (higher during the day than at night), and contamination with secretions from the urinary tract etc; hence new occurrences should be confirmed using an early morning urine. | | | | | |
| No reference range is quoted for urine albumin concentration as it varies with urine dilution. Results are always reported as albumin/creatinine ratio (ACR). | | | | | |
| ACR has a long established role in diabetes, to detect microalbuminuria (ACR persistently high but below 30mg/mmol). | | | | | |
| Amino acids | Random urine - plain | See report | GOS | 2 weeks | Send fresh urine sample promptly to lab |
| Urine amino acid analysis should only be requested for the identification of renal amino acid transport disorders (e.g., cystinuria, lysinuric protein intolerance or hartnup disease) or as a marker of renal tubular function. The test is also indicated in the investigation of hypophosphatasia and prolidase deficiency. | | | | | |
| For the majority of amino acids renal tubular (re) absorption should be >95%. | | | | | |
| Please note that the plasma is the sample type of choice for the investigation of most inherited disorders of amino acid metabolism. | | | | | |
| Amylase | 24 hr urine collection - plain | 0-1100 IU/24hrs | | <1 day | |
| Arsenic | Random urine - plain | See report | HALO | 2 -3 weeks | |

| Test | Sample | Adult Reference range | *If referred site code | Turnaround time | Comments |
|---|---|--|----------------------------|--------------------|---|
| Bence Jones protein (BJP) | Random urine - plain | See report | HALO | <7 days | |
| Calcium | 24hr collection into 25 mL 5M HCL | 2.5-7.5 mmol/24hrs | | <1 day | |
| Chloride | 24 hr urine collection - plain | 110-250 mmol/24hr | | <1 day | |
| Citrate | 24hr collection into 25 mL 5M HCL | Male 0.6-4.8 mmol/24hr Female 1.3- 6.0mmol/24hr | HALO | 2 - 3 weeks | |
| Copper | 24 hr urine collection - plain | 0-1.2 μ mol/L | HALO | < 2 weeks | |
| Cortisol (urine free Cortisol, UFC) | 24 hr urine – plain | 50-270nmol /24hr | HALO | <2 weeks | |
| Creatinine | 24 hr urine collection – plain, or random urine | See comments | | < 3 days | Reference ranges: 6-13 mmol/24h (female) 9-19 mmol/24h (male) |
| Urine creatinine is rarely required on timed collections, except when used as a measure of dialysis adequacy. It is mainly used as a component of other urine tests, to compensate for variation in urine dilution (eg. ACR and PCR). | | | | | |
| Creatinine Clearance | | 60-120ml/min | ADI | < 3 days | |
| Cobalt | Random urine - plain | Interpretation with report | CHX | < 2 weeks | |
| Cystine | 24hr collection into 25 mL 5M HCL | 0-600 μ mol/ 24hr | HALO | 2 - 3 weeks | |
| Diuretic Screen | Random urine - plain | See report | City Hospital, B'ham | 1 week | |

| Test | Sample | Adult Reference range | *If referred site code | Turnaround time | Comments |
|--|--|-----------------------------|---------------------------|--------------------|--|
| Drugs of Abuse screen | Random urine - plain | Not detected | SAN | 2 -3 weeks | Includes Creatinine, amphetamines, barbiturates, benzodiazepines, THC, cocaine, methadone & opiates. |
| FIT (Faecal immunoturbidimmetry) | Sample kit provided for faecal sample | See report | HALO | 5 days | |
| Glycolate | 24hr collection into 25 mL 5M HCL | 4-36 μmol/mmol creat | UCLH | 2 -3 weeks | |
| HVA (homovanillic acid) | See VMA | | | | |
| Iodine | 24 hr urine collection - plain | | CHX | 1 month | |
| Laxatives | Random urine - plain | See report | HALO | 2 weeks | Discuss case with laboratory consultant first |
| Mercury | 24 hr urine collection - plain | <5 nmol/mmol creat | HALO | <2 weeks | Use for exposure to mercury vapour or inorganic mercury. Use blood test for organic mercury |
| Metabolic stone screen (urine) | 24 hr urine collection - plain PLUS 24hr collection into 25 mL 5M HCL | See report | HALO | 2-3 weeks | Profile includes: sodium, potassium, calcium, citrate, cystine, oxalate, & urate |
| Metadrenalines (metadrenaline, normetadrenaline & 3-methoxy tyramine) | 24hr collection into 25 mL 5M HCL | See report | STH | <2 weeks | |
| Urine metadrenalines test is the first-line investigation used in the diagnosis of suspected phaeochromocytoma. | | | | | |
| Note that severe stress (MI, hypoglycaemia, surgery) and some drugs can cause physiological increase in some or all of the metadrenalines (eg. all dopaminergic drugs such as L-dopa and methyldopa, doxazosin, tricyclic anti-depressants, phenoxybenzamine, MAO inhibitors, methylphenidate, amphetamine). Consideration should be given to stopping these drugs before advising the patient to do the urine collection. | | | | | |

| Test | Sample | Adult Reference range | *If referred site code | Turnaround time | Comments |
|--|--------------------------------------|-----------------------------|---------------------------|--------------------|---|
| Metadrenalines, random urine | Random urine - plain | See report | STH | <2 weeks | |
| <p>This test must only be used in children who are not capable of accurately collecting a 24h urine specimen. It is the first-line investigation in the diagnosis of suspected phaeochromocytoma; for suspected neuroblastoma in children, request random urine VMA instead.</p> <p>Samples must be fresh and handed directly to a member of laboratory staff for acidification.</p> <p>Do not request this test in adults: instead use 24h urine metadrenalines. This is because reference ranges and interpretation of 24h urines are more robust than those for random urines.</p> | | | | | |
| <p>Microscopy (Sedimax analyser)</p> <p>Random urine- can be collected at any time but first thing in the morning is the recommended optimal time of collection</p> <p>Sterile Boric acid container preferred as preserves sample for longer but sterile plain also accepted.</p> <p>A minimum of 2.5ml of sample is required for microscopy however a dilution can be performed with as little as 0.5ml of sample. If a sample needs to be sent for culture then more will be required.</p> <p>Analysis occurs same day in-house between 9am and 5pm Monday to Friday.</p> <p>Out of hours samples are sent directly to CUH for analysis via regular Citysprint shuttle run to microbiology department.</p> | | | | | |
| Mucopoly- saccarides | Random urine - plain | See report | GOS | 4 weeks | |
| Magnesium | 24 hr urine collection - plain | | | <3 days | Discuss case with laboratory consultant first |
| Magnesium | Random urine – plain | | | < 1 day | May be used to determine cause of hypomagnesaemia. |
| Mercury | Random urine - plain | See report | | | 24h urine may be used if preferred |
| <p>This test should be used only for cases of suspected poisoning with metallic mercury or inorganic mercury compounds.</p> <p>Do not use in relation to dental amalgam fillings. There is no convincing evidence that these cause any adverse health effects.</p> | | | | | |
| N-Telopeptide Crosslinks NTX | Random urine - plain | | HALO | 3 weeks | |
| Osmolality | Random urine - plain | 40-1400 mOsmol/kg | | <1 day | For water deprivation test, consult laboratory consultant first |
| Organic acids | Random urine - plain | See report | HALO | 2-4 weeks | Fresh sample required |

| Test | Sample | Adult Reference range | *If referred site code | Turnaround time | Comments |
|---|--|-----------------------------|---------------------------|--------------------|--|
| Urine organic acid analysis is a core metabolic test and is useful aid for the diagnosis of organic acidurias and a number of inherited metabolic disorders including those of amino acid metabolism (e.g., urea cycle defects, Maple Syrup Urine Disease). | | | | | |
| Oxalate | 24hr collection into 25 mL 5M HCL | 100-460 µmol/24hr | HALO | 2 -3 weeks | |
| pH Urine | Random urine collected in a universal container. | | | < 1 day | pH should be analysed within an hour of sample collection |
| Phosphate | 24hr collection into 25 mL 5M HCL | 15-50 mmol/24hr | | <3 days | |
| Porphobilinogen (PBG) | Random urine - plain Exclude light | See report | HALO | <2 weeks | Can be expedited if acute situation. |
| Purines and pyrimidines | Random urine - plain | See report | | | Send fresh sample promptly to lab |
| Porphyrins | Random urine - plain Exclude light | See report | HALO | <2 weeks | |
| Steroid profile | Random urine - plain | See report | UCLH | 2 -3 weeks | |
| Sulphite | Random urine - plain | See report | | 2-3weeks | Send fresh sample promptly to lab |
| Sulphocysteine | Random urine - plain | See report | UCLH | 2-3weeks | Send fresh sample promptly to lab |
| This test is used in the diagnosis of molybdenum cofactor deficiency and isolated sulphite oxidase deficiency. | | | | | |
| Potassium | 24 hr urine collection - plain | 35-100 mmol/24hrs | HALO | <3 days | |

| Test | Sample | Adult Reference range | *If referred site code | Turnaround time | Comments |
|--|--|---|---------------------------|--------------------|--|
| Protein | 24 hr urine collection - plain OR random urine - plain | <0.14g/L OR reported as PCR, see comments | | <1 day | Reference range for PCR: < 15mg/mmol creatinine |
| Urine protein is used in the diagnosis and monitoring of kidney disease and dialysis efficiency. It is also used in the diagnosis of hypertensive disorders of pregnancy. | | | | | |
| No reference range is quoted for urine protein concentration as it varies with urine dilution. Results are always reported as protein/creatinine ratio (PCR). | | | | | |
| In general, random urine PCR is the recommended test, especially when used in pregnancy. | | | | | |
| Sodium | 24 hr urine collection - plain | 110-240 mmol/24hrs | HALO | <3 days | |
| Urate | 24 hr urine collection - plain | 1.5-4.5 mmol/24hrs | HALO | <3 days | |
| Urea | 24 hr urine collection - plain OR random urine - plain | 428-714 mmol/24h | | <3 days | |
| Urine urea is mainly used for monitoring dialysis efficiency | | | | | |
| VMA (vanillylmandelic acid) | Random urine - plain | | ADI | <4 days | Assay is routinely run on Tuesdays and Thursdays |
| VMA test is performed with HVA (homovanillic acid). Together they are used in the diagnosis and monitoring of neuroblastoma. Urgent diagnostic tests can be reported within 24h if they are Consultant requests and agreed in advance. | | | | | |
| This test is about 90% sensitive for neuroblastoma; a normal result does not entirely exclude it. Several drugs can interfere, especially L-dopa. | | | | | |
| Samples must be fresh and handed directly to a member of laboratory staff for acidification. | | | | | |
| For suspected phaeochromocytoma in children, request plasma metadrenalin instead. | | | | | |

Specialist Metabolic Blood Tests

Indicative turnaround times are given for non-urgent situations. It may be possible to expedite results by discussing the clinical situation with the Laboratory Consultant staff. If a required test is not listed here, please discuss with the Laboratory Consultant staff.

| Test | Sample | Comments | Turnround |
|---|---|---|-----------|
| Acylcarnitines | 4 dried blood spots (Guthrie card) | Clearly write "acyl carnitines" on the card. Allow to air-dry, then place in glassine sleeve and send to Laboratory. Do not post to Newborn Screening Laboratory. | <2 weeks |
| Amino acids <i>includes homocysteine, glycine, serine</i> | Heparin (Green) 1mL | Avoid haemolysis. Send sample to Laboratory within 30 minutes. | 2 weeks |
| Ammonia | EDTA (Lavender) 1mL | Inform Laboratory. Avoid haemolysis. Take to laboratory immediately. Within 15 minutes. | 3 hours |
| β -hydroxybutyrate | Heparin (Green) 0.5mL | Send sample to Laboratory within 30 minutes. Send sample for glucose at same time. | 1 week |
| Biotinidase | Heparin (Green) 0.5mL | Avoid haemolysis. Send sample to Laboratory within 30 minutes. | <2 weeks |
| C-peptide | Clotted (red or Yellow) preferred Heparin (Green) also acceptable 0.5mL | Send sample to Laboratory within 30 minutes. Send sample for glucose at same time. Please note adult and paediatric samples are sent to differing locations | 1 weeks |
| 7-dehydro- cholesterol | Heparin (Green) 1mL | Requests are easily misinterpreted as cholesterol. To avoid this, please alert Laboratory before sending | 8 weeks |
| Free fatty acids | Heparin (Green) 0.5mL | Send sample to Laboratory within 30 minutes. Send sample for glucose at same time. | 1-2 weeks |
| β -galactosidase | EDTA (Lavender) 5mL | Send straight to the lab. For Fabry disease in males and females. Confirmation testing for females follows the rare and inherited diseases - molecular genetics pathway (GEN1 on ICE) | 5 weeks |
| β -glucuronidase (acid maltase) | EDTA (Lavender) 2mL | For Pompe disease (Glycogen Storage Disease Type II) | 5 weeks |

| Test | Sample | Comments | Turnround |
|--|---|---|------------|
| Gal-1-PUT | Heparin (Green) 2mL | Contact Laboratory to arrange: needs rapid transport to another Laboratory. Do not collect if transfusion in previous 6 weeks | 4 weeks |
| Insulin | Clotted (red or Yellow) preferred Heparin (Green) also acceptable 0.5mL | Avoid haemolysis. Send sample to Laboratory within 30 minutes. Send sample for glucose at same time. | 2 weeks |
| Lactate | Fluoride (grey) 1mL | Request glucose on same sample | 3 hours |
| Transferrin glycoforms | Heparin (Green) or clotted (red or Yellow) 0.5mL | | 10 days |
| Ubiquinone | EDTA (Lavender) 5-10mL | Contact Laboratory to arrange: needs rapid transport to another Laboratory | 4 weeks |
| Vacuolated lymphocytes | EDTA (Lavender) 2mL | Contact Laboratory to arrange: needs rapid transport to another Laboratory. For lysosomal storage disorders | Not stated |
| VLCFA (very long chain fatty acids) <i>Includes phytanate and pristanate</i> | Heparin (Green) 1mL | Fasting sample preferred. Send sample to Laboratory within 30 minutes. | 2-4 weeks |
| White cell enzymes | Heparin (Green) 5-10mL | Contact Laboratory to arrange: needs rapid transport to another Laboratory. Full clinical details must be given. | 6-8 weeks |

Specialist metabolic Urine investigations

Indicative turnaround times are given for non-urgent situations. It may be possible to expedite results by discussing the clinical situation with the Laboratory Consultant staff. If a required test is not listed here, please discuss with the laboratory consultant staff.

| Test | Sample | Comments | Turnround |
|--|--------------------------------------|--|------------|
| Urine amino acids | Random urine, plain container 5mL | Send fresh sample promptly to Laboratory | <2 weeks |
| Urine amino acid analysis should only be requested for the identification of renal amino acid transport disorders (e.g., cystinuria, lysinuric protein intolerance or hartnup disease) or as a marker of renal tubular function. The test is also indicated in the investigation of hypophosphatasia and prolidase deficiency. | | | |
| For the majority of amino acids renal tubular (re) absorption should be >95%. | | | |
| Please note that the plasma is the sample type of choice for the investigation of most inherited disorders of amino acid metabolism. | | | |
| Urine mucopolysaccharides (glycosaminoglycans) | Random urine, plain container 5mL | Send fresh sample promptly to Laboratory | 6-8 weeks |
| Urine organic acids | Random urine, plain container 5mL | Send fresh sample promptly to Laboratory | 3 weeks |
| Urine purines and pyrimidines | Random urine, plain container 5mL | Send fresh sample promptly to Laboratory | Not stated |

| | | | |
|----------------------|-----------------------------------|--|---------|
| Urine sulphite | Random urine, plain container 5mL | Send fresh sample promptly to Laboratory | 1 week |
| Urine sulphocysteine | Random urine, plain container 5mL | Send fresh sample promptly to Laboratory | 3 weeks |

Specialist metabolic CSF tests

Indicative turnaround times are given for non-urgent situations. It may be possible to expedite results by discussing the clinical situation with the Laboratory Consultant staff. If a required test is not listed here, please discuss with the Laboratory Consultant staff.

| Test | Sample | Comments | Turnaround |
|--|-----------------------|---|------------|
| CSF amino acids <i>includes glycine, serine</i> | Plain container 0.2mL | Avoid contamination with blood. Send fresh sample promptly to Laboratory. Send plasma for amino acids at same time. | 2 weeks |
| CSF lactate | Fluoride (grey) | Request glucose on same sample. Send blood sample for glucose and lactate at same time | 3 hours |
| CSF neurotransmitters <i>includes monoamine metabolites, 5-methyl tetrahydrofolate, pterins, pyridoxal phosphate</i> | | Not available locally | |

Requesting genetic tests in East & North Herts NHS Trust

The NHS Genomic Medicine Service has improved access to genomic testing and standardised requesting processes through a network of Genomics Hubs. Funding for our patients is provided centrally to our local Genomics Hub Laboratory (East Genomic Laboratory Hub, located in Cambridge) and all blood samples for genomics tests must be sent there. Requests must be made in ICE and the user will be asked the appropriate questions for the CUH Genomics hub to use (See below).

Requesters are advised to familiarise themselves with the “Rare and inherited disease eligibility criteria” (visit <https://www.england.nhs.uk/publication/national-genomic-test-directories/>) before making requests.

The reporting time guideline for most of this service is 6 weeks (42 days). For further guidance, see <https://www.eastgenomics.nhs.uk/about-us/quality/nhse-test-directory-turnaround-times/>

Requests from this Trust must be made using our ICE Ordercomms system:

- If you have been using paper genomics request forms published by other laboratories (eg. Great Ormond Street, Royal Brompton, etc), please stop this practice as it causes confusion over the location of the requester.
- In the search panel on ICE, type in “rare”. For Molecular Genetics tests (DNA tests, and microarray – EDTA blood sample) choose the option ending in **Mol**. For Cytogenetics tests (chromosome tests such as karyotype, FISH, X-inactivation testing - heparin blood sample) choose the option ending in **Cyto**.
- Confirm that you have discussed genomic testing with the patient and retained a record in the notes (tick **yes** or **no**)
- State exactly the test required (if possible give the relevant R code from “Rare and inherited disease eligibility criteria”), and give a full description of the clinical indication including results of related tests, index case identifiers

etc. This is a free-text box that will expand as you type. Pedigree diagrams cannot be entered on ICE - please attach on a separate piece of paper if relevant.

- Select the type of test required from the drop-down menu (diagnostic, predictive, carrier, or DNA storage only).
- Enter your email address so that the Genomics Hub can contact you for further information if required.
- Click OK.
- There are optional links that you can follow from ICE to the National Genomic Test Directory for Rare and Inherited Disease, or to the genomics hub's guidance on consent.
- Results will be reported on ICE.

Requests by Genetic Counsellors made on behalf of this Trust:

If you are requesting from an off-site location, you may not have access to ICE requesting. In this case, please continue to use your previous paper request forms, or the forms from the East Genomics Hub, observing the following:

- Type your entries if possible, or take great care to ensure full legibility.
- State the blood tube type required (EDTA or heparin, or both), if that is not otherwise clearly on the form, otherwise the Phlebotomist may not be able to collect the blood for you.
- State the R-code as well as other clinical information as this speeds the flow through the Genomics Hub Lab.
- Be sure to state your name and email address, and make it clear that you are operating from either the Lister Hospital OPD, North-West Thames Genetics Service, or Northwick Park Hospital. Results will be reported on ICE.

Questions or problems:

If you experience any problems or have any queries or suggestions regarding the local part of the requesting process, please contact either: angelawoods@nhs.net or enhtr.iceocs@nhs.net.

Haematology

Haematology is part of Blood Sciences department.

Opening hours

Monday to Friday: 09:00 - 17:00.

At all other times contact the Biomedical Scientist out of hours for urgent work only. Bleep 1005

Key contacts

General Queries: 01438 284961

Consultant haematologists

Dr J Hanslip judith.hanslip@nhs.net 01438 285375 (Ext 5375)

Dr Xenofon Papanikolaou xenofon.papanikolaou@nhs.net 01438 284159 (Ext4159)

Dr Muhammad Hasan muhammad.hasan1@nhs.net 01438 281610 (Ext1610)

Dr Aushna Rasool Aushna.rasool@nhs.net

Dr Ama Batool asma.batool@nhs.net

Essential Services Laboratory Manager

Kate Barrett

E: kate.barrett@hslpathology.com

Results

The Sunquest ICE system should be used to look for results. In the event that this is not possible Haematology should be contacted on 01438 285461 (Ext 5461).

Critical results

These apply to acute work only, Results outside the limits below will be telephoned urgently to the requesting clinician.

Results from a positive malarial parasite screen.

ALL SIGNIFICANTLY ABNORMAL RESULTS, whether urgent or not must be telephoned to the requesting clinician unless previous results, already reported, have been similar (+/- 10 %).

| Test | Units | Critical Low | Critical High |
|--------------------|-------------------------|--|---------------|
| WBC | $\times 10^9$ L | < 1.0 | > 50.0 |
| Neutrophils | $\times 10^9$ L | < 0.5 | > 50.0 |
| Lymphocytes | $\times 10^9$ L (adult) | | > 50.0 |
| Hb | g/L | <70 Normchromic, or <50 Macro or Micro-Cytic | > 190 |
| Platelets | $\times 10^9$ L | <30 * | >1000 |
| INR (warfarin pts) | Ratio | | >6.0 |
| APTT Ratio | Ratio | | >3.0 |

| | | | |
|------------|---------|------|--|
| PT | Seconds | | >20 |
| APTT | Seconds | | >60 |
| Fibrinogen | g/L | <1.2 | |
| DDimer | | | >3000ng/ml |
| ESR | mm/hour | | >80 (with symptoms such as headache, giant cell arteritis or temporal arteritis) |

Specimens and tests - tests marked * in the table below have an expected completion time of <1 hour from receipt in lab if requested urgently from A/E.

Sample volumes 4ml EDTA or 5ml Citrate unless otherwise stated.

Please do not collect EDTA samples in 6ml or 10ml unless stated otherwise.

| Test | Sample | Reference range | Comments/referral site | Turnaround |
|---|---------------------|--|--|--|
| ANEP | EDTA 4ml (Lavender) | | Haemoglobinopathy screens performed as part of the Antenatal haemoglobinopathy screening programme must be accompanied by a completed Family Origin Questionnaire. Abnormals are referred to the HALO | <3 days |
| Full Blood Count * | EDTA (Lavender) | WBC 4-11.0 10^9 /L HB Female 115-160 g/L Male 130-170 g/L Platelets 150-400 10^9 /L | | <8 hours |
| Haematologist Film Comment | EDTA (Lavender) | | | 24 - 48 hours Urgently as required. |
| Erythrocyte Sedimentation Rate (ESR) *Urgent TAT 2hrs (analysis takes 1hr) | EDTA 4ml (Lavender) | Male <50 yr 1-10 50-60 yr 1-12 60-70 yr 1-14 >70 yr 1-30 Female <50 yr 1-19 50-60 yr 1-19 60-70 yr 1-20 >70 yr 1-35 | Exact quantity of blood required – allow blood to fill vacuum. | <8 hours |
| Reticulocyte count | EDTA 4ml (Lavender) | 30-90 10^9 /L | | <8 hours |

| | | | | |
|---|--|---|--|-----------|
| Infectious Mononucleosis Test (glandular fever) | EDTA 4ml (Lavender) and a clotted sample | | | <24 hours |
| Malaria Screen * Includes blood film and immuno test | EDTA 4ml (Lavender) | If both results are negative but malaria suspected clinically, sample should be repeated 2-3 times at 8-12h intervals. (Note that immuno test may not detect P. knowlesi) | | <4 hrs |
| Haemoglobinopathy Screen | EDTA 4ml (Lavender) | Haemoglobinopathy screens performed as part of the Antenatal haemoglobinopathy screening programme must be accompanied by a completed Family Origin Questionnaire. Abnormals are referred to Addenbrookes Hospital | | <3 days |
| Sickle screen* | EDTA 4ml (Lavender) | | Confirmed by Haemoglobinopathy screen. For confirmation please request haemoglobinopathy screen on ICE. HBEP (non antenatal) or ANEP (antenatal) | <4 hours |
| Clotting screen PT APTT Fibrinogen (Fib) | Citrate (Blue) PT 9-12 secs APTT 22-28 secs Fib 2-4g/L | | PT, APTT and Clauss fibrinogen performed on all coagulation screens. Fill sample tube to mark. | < 8 hours |
| INR (oral anticoagulant control)* | Citrate (Blue) | Patient specific therapeutic range | Fill sample tube to mark. | < 8 hours |
| D-Dimer* | Citrate (Blue) | 0-500 ng/ml | Fill sample tube to mark. | < 4 hours |
| 'Thrombophilia' screens. Lupus Anticoagulant | EDTA 4ml (Lavender x4) | Refer to report for result interpretation Please see guidelines on appropriate testing. | | 2-3 weeks |

| | | | | |
|---|--|--|---|---------|
| | | | Sent to Royal Free Hospital | |
| Haematinics, B12, serum folate,(FOLF/FOLS) ferritin (FER) | EDTA (Lavender) and a clotted sample (for serum folate a 12hr fast is recommended, but not essential | B12 200-770ng/L Serum Folate* >4.5 µg/L <3 µg/L is indicative of folate deficiency FER Male 30-400-ug/L Female 13-150ug/L | B12 guidance no sample should be collected from a patient who has received Vit B12 injection therapy within the past week *A folate rich meal taken 3hours before venesection may increase serum folate concentration Age related references reportable. | <3 days |
| Factor V Leiden | EDTA 4ml (Lavender) | | Referred to the HALO if not part of a thrombophilia screen. | 7 days |
| Coagulation Factor Assays | Citrate (Blue) | See report for interpretation | Referred to Royal Free Hospital | <72 hrs |
| Anti-Xa assay | Citrate (Blue) | | Requires prior discussion with Haematology Consultant | |
| Apixaban Anti Xa | Citrate (Blue) | | Referred Royal Free Hospital | |
| Edoxaban Anti Xa | Citrate (Blue) | | Referred Royal Free Hospital | |
| Rivaroxaban Anti Xa | Citrate (Blue) | | Referred Royal Free Hospital | |
| Von Willebrands factor | Citrate (Blue) | | Referred Royal Free Hospital | 3 days |
| G-6-PD screen | EDTA 4ml (Lavender) | | If deficient sent to HSL | <48 hrs |
| Haemochromatosis | Crossmatch tube | | Referred to HALO | 6 weeks |
| Haemosiderin | Urine | | | 2 days |

| | | | | |
|--|---------------------------------------|---|--|---------|
| | | | | |
| SPECIAL HAEMATOLOGY Bone Marrow Aspirates and Trehpine Biopsies Schilling tests | Discuss with Consultant Haematologist | | | |
| Immunophenotyping Molecular tests Jak 2 | Crossmatch tube x2 | Refer to report for result interpretation | Referred to SIHMDS/UCLH | 2 Weeks |
| BCR/ABL | Crossmatch tube x2 | Refer to report for result interpretation | Two sample tubes Referred to SIHMDS/UCLH for screening. | 2 weeks |

The laboratory must be notified by telephone of all cases that are deemed clinically urgent.

Blood transfusion

Blood Transfusion is part of Blood Sciences department.

General enquiries Lister 01438 285245

Opening hours

Lister Monday to Friday: 9:00 - 17:00

At all other times contact the Biomedical Scientist on call for urgent work - Bleep 1005

Key contacts

Consultant Haematologist & Clinical lead

Dr M Xenofon Papanikolaou xenofon.papanikolaou@nhs.net 01438 284146 (Ext 4146) Bleep 5922

Essential Services Laboratory Manager

Kate Barrett kate.barrett@hslpathology.com

Transfusion Lead Biomedical Scientist

Jane Tidman - j.tidman@nhs.net 01438 2844295 (EXT 4295)

Specialist Practitioners of Transfusion

Mrs S Needham sheila.needham@nhs.net 01438 288016 (Ext 8016 or 8017)

Mrs K Baylis karen.baylis@nhs.net 01438 288017 (Ext 8016 or 8017)

Mrs J Edmonds julie.edmonds@nhs.net 01438 288016 (Ext 8016 or 8017)

Specimens and tests

All staff involved with the collection of blood samples for the transfusion laboratory or with the transportation /administration of blood products must undergo training and competency assessment in order to comply with the requirements of the National Patient Safety Advice SPN14.

All samples (4.9ml EDTA tubes) and request forms for the blood transfusion laboratory must be labelled with four points of identification:

- NHS Number
- Surname
- First name
- Date of birth
- Hospital number

Details on sample tubes must be hand written and the tube signed by the venesector. All details on the tube must be identical to those on the form and corroborate with the patients wristband and clinical notes. Any sample/request form that has incomplete or discrepant labelling will not be processed.

Blood Group and Antibody Screen (Antenatal)

Clinically significant antibodies in pregnancy are monitored in conjunction with the National Blood Service and are brought to the attention of the Consultant Haematologist for appropriate action.

Blood group and antibody screen (group and save)

This Trust operates a two sample policy. Before cross-matched blood can be provided there has to be at least two group results available for the patient. These samples must be taken on two separate occasions at least 15 minutes apart. Timely provision of compatible blood can be expedited much more effectively when a request for a group and antibody screen is sent with sufficient notice to allow the identification of any clinically significant antibodies that may be present. Where any clinically significant antibodies are detected a delay in the provision of compatible blood may be unavoidable. Turnaround time for routine group and save is 24 hours

Regularly transfused patients

Many Regularly Transfused Patients have complex special requirements that necessitate their blood being prepared at Colindale Blood Transfusion Centre. Once Colindale has issued compatible blood it is only valid for use for 72 hours from the time the sample was taken. Others require blood to be ordered in specifically to meet their needs. In order to ensure we have blood available for these patients we need to receive the requests and samples for all regularly transfused patients 48 hours prior to the scheduled transfusion time.

Cross matching

Compatible blood is provided for all patients needing transfusion. For routine blood transfusion the same day, cross-match samples must be received in the laboratory by 13:00 hours. In the case of elective surgery please refer to the Maximum Surgical Blood Ordering Schedule when deciding which request to make for a particular operation. The Schedule is included in the Junior Doctors' induction pack and available on the Trust intranet. It is essential to provide the nature, date and time of the proposed surgery on the request form. Please discuss variations from the schedule with the laboratory. A blood group and antibody screen should be ordered at the out-patient appointment or at the pre-admission clinic visit. There needs to be a repeat sample taken within the week of the scheduled surgery, with any request for crossmatching, to ensure a valid sample is available. Where blood provided to cover an operation is not used, it will be reclaimed the following morning without notice unless prior arrangements have been made with the Transfusion Laboratory.

Requests for the rapid provision of blood must be telephoned to the laboratory. If the criteria for Electronic Issue are met blood can be made available in 10 minutes providing there is a valid group and save. When serological cross-match is required the minimum turn around time is 1 hour depending on any antibodies detected, with a valid group and save already in the laboratory.

Blood transfusion policy

A comprehensive set of Blood Transfusion Policies are available in all clinical areas and on the Hospital Intranet / Knowledge Centre. It covers all aspects of requesting and the administration of blood and blood components, and is applicable to all hospitals in East and North Herts NHS Trust.

Fresh Frozen Plasma (FFP), Cryoprecipitate and Platelet concentrates can be ordered directly from the laboratory. Routine orders for Platelets must be made before 10.30am. Clotting Factor concentrates can be ordered after discussion with a Consultant Haematologist.

Advice and interpretation of results

The Consultant Haematologists are available to advise users on how to use the Laboratory most effectively. They are happy to discuss the best way to investigate a specific clinical problem, to comment on abnormal results and to make suggestions regarding further investigation, if required.

Comments may be sent out, as an addendum to the Laboratory report but in complex or urgent situations a telephone call will be initiated by either the Consultant Haematologist or the user.

BT Referral Tests

| Description | Form | Destination | Codes |
|---|------|---------------|-------|
| Adult Autoimmune Neutropenia | 3E | Filton | AAIN |
| Antibody Identification | 1A | RCI Colindale | ABID |
| Anti A ₁ and B Titre | 1A | RCI Colindale | ABTR |
| Antibody Titre | 1A | RCI Colindale | TITR |
| Autoimmune Thrombocytopenia | 3D | Filton | AIT |
| Blood Group Confirmation | 1A | RCI Colindale | BGCN |
| Crossmatch Components | 1A | RCI Colindale | XMC |
| Drug Related Thrombocytopenia | 3D | Filton | DRT |
| Drug Related Neutropenia | 3E | Filton | DREN |
| Fetal/ Neonatal Alloimmune Thrombocytopenia | 3D | Filton | NAIT |
| Flow Cytometry | 1A | RCI Colindale | FCC |
| Heparin Induced PLT Antibodies | 3D | Filton | HIPA |
| HFE/ Haemochromotosis | 3F | H&I Colindale | HFE |
| HLA Testing for Narcolepsy | 3F | H&I Colindale | NAR |
| HLA ABC Typing (HLA Type) | 3F | H&I Colindale | ABC |
| HLA B27 | 3F | H&I Colindale | B27 |
| HLA B*57:01 | 3F | H&I Colindale | 5701 |
| HLA DR Typing | 3F | H&I Colindale | DR |
| HPA Type | 3D | H&I Colindale | HPAT |
| HNA Type | 3E | H&I Colindale | HNAT |
| HLA Antibody Screen | 3F | H&I Colindale | HLAS |
| HPA Antibody Screen | 3D | H&I Colindale | HPAS |
| HNA Antibody Screen | 3E | H&I Colindale | HNAS |
| Investigation of Platelet Refractoriness | 3D | H&I Colindale | IPR |
| Infant Autoimmune Neutropenia | 3E | Filton | IAIN |
| Neonatal Alloimmune Neutropenia | 3E | Filton | NAIN |
| Neutrophil Antibodies | 3E | Filton | NEUA |
| Rh D confirmation | 1A | RCI Colindale | DVAR |

| | | | |
|--|-----------|---------------|------|
| Post Transfusion Purpura | 3A | Filton | PTP |
| Severe febrile non-haemolytic transfusion reaction | 3A | H&I Colindale | SFTR |
| Transfusion-Related Acute Lung Injury | 3A | H&I Colindale | TRAL |
| Transfusion-associated graft versus host disease | 3A | H&I Colindale | TAGD |
| Foetal RHD Screen | FRH5197/3 | Filton | FDNA |

FOR REFERENCE ONLY

Immunology

Most Immunology requests are tested HSL hub laboratory at 1 Mabledon place. For further information please see the HSL user guide:

Key contacts

Consultant immunologist:

Dr Scott Pereira

scott.pereira@nhs.net

07798914734

Halo Immunology urgent contact information: **Urgent ANCAs (MPO, PR3, GBM)**

Allergy bench: 0203 908 1471

Kushen Ramessur Head of Immunology: 077 0360 9084

Our opening hours are Monday - Friday 8:00 - 17:30
Saturday 9:00 - 16:30
Sunday Closed

Results

All results are available on ICE system

Immunology sample requirements

Serum Samples

Most tests are performed on serum separated at room temperature. The exceptions are listed below. For serum, blood should be collected in a plain yellow top tube with no anticoagulant.

Allergy testing

The specific allergen required must be clearly stated.

Where mixes of multiple varieties are available, they will be used unless a specific allergen is requested e.g.

Grass mix performed, unless 'Timothy grass' is specified.

If the request is unclear Immunology at HALO may perform general food allergy testing, although it cannot accept responsibility for any choices made.

A request for Aspergillus will receive IgE, unless IgG Aspergillus is clearly stated.

If total IgE measurement is required, it must be clearly requested.

Lymphocyte Phenotyping Studies

These are performed on a 5 ml EDTA (Lavender) sample in HALO Haematology

Quantiferon TB Gold (Interferon gamma release assay-IGRA)

Adults and children: 1ml of blood must be drawn directly into each of the four QuantiFERON-GOLD Plus tubes in order, which must be all labelled with appropriate patient identifying information.

(1) Nil control (GREY TOP)

(2) TB 1 Antigen (GREEN TOP)

(3) TB Antigen 2 (YELLOW TOP)

(4) Mitogen Control (PURPLE TOP)

(Collection tubes available from Phlebotomy and directly from the Immunology laboratory)

Samples must be incubated at 37o c within 16 hrs of collection at HALO, and must be kept at room temperature prior to this.

Samples must be received into the laboratory by 6pm Friday for processing.

Samples received after this time will not be tested.

Cytokine studies

Please send from Patient and from a healthy control:

5-10 ml Li-heparin blood (3 ml from very small children)

2.7 ml Edta-blood

1-2 ml Serum/clotted (not for very small children)

Cryoglobulins

If cryoglobulins are suspected in autoimmune rheumatoid disorders, a 10 ml blood sample should be

collected in a *pre-warmed* plain white Monovette tube and placed immediately into a vacuum flask

Contact the laboratory for assistance.

Requestors should be aware of the testing protocol before contacting laboratory.

Immunoglobulins

Routine tests for total serum IgG, IgA and IgM are performed in the Clinical Biochemistry. Tests for IgG subclasses, IgE, and very low immuno-globulins in hypogammaglobulinaemia are performed in the Clinical Immunology laboratory.

Microbiology

These notes are provided for clinical staff using the microbiology laboratory; they are not intended to be a complete or authoritative document but merely a guide to some of the services available. If you need further information about specimens, availability or suitability of tests, interpretation of results, or any other matter relating to the microbiology service, phone the department - staff will be pleased to help.

All microbiology testing is performed at the HSL hub laboratory at 1 Mabledon place. For further information please see the HSL user guide: hslpathology.com/media/eohdxhdv/tap5622_hsl_user_guide_2025_v1.pdf

Key contacts

Lead Clinician

Dr Eleni Mavrogiorgou
elenimavrogiorgou@nhs.net
Direct line 01438 284288 (Ext 4288)
Secretary Direct line 01438 284150 | Ext. 4150

Consultant Microbiologists:

Dr Eleni Mavrogiorgou
Direct line 01438 284288 (Ext 4288)
Secretary Direct line 01438 284150 | Ext. 4150

Dr Saba Qaiser
Direct line 01438 284578 (Ext 4578)
Secretary Direct line 01438 284150 | Ext. 4150

Dr Zoi Foka
Direct line 01438 284288 (Ext 4288)
Secretary Direct line 01438 284150 | Ext. 4150

Laboratory Manager

Kate Barrett kate.barrett@hslpathology.com

Infection Control

infectionprevention.enh-tr@nhs.net

01438 285383

Contact via Alertive

Clinical Microbiology Advice Line

microbiologyadvice.enh-tr@nhs.net

Monday-Friday 9am-5pm 07500 975834

Out of hours: Microbiology Consultant on call via switchboard.

Microbiology referral checklist

The on-call microbiology consultant can be contacted via switchboard.

Please know your patient well and seek senior advice prior to contacting microbiology.

Before contacting microbiology, please ensure you have the following information at hand in addition to your query:

- Patient name
- Age
- NHS number
- Ward
- Consultant

- Allergies (including nature of allergy)
- Any relevant past medical history (including any previous relevant infections)
- Day of admission
- Presenting complain/ infection at hand
- Current antibiotics
- Previous antibiotics (for this infection)
- Blood tests and blood cultures
- Any relevant imaging results (e.g. cardiac echo)
- Renal function/ modality
- Discharge plan if relevant
- Name and bleep number

Specimens should be left at pathology reception and these will be transported to HALO. During normal working hours you must ensure the laboratory is informed by telephone of any urgent specimens and that the specimens are labelled 'URGENT'.

Without the full information detailed above, it is impossible to examine a specimen or to accurately report it.
For minimum acceptance criteria for making pathology requests see Pathology Appendix 2- Trust Policy for Pathology Sample Collection.

All specimens must be labelled using the pull off labels from the Sunquest ICE request form, and the date, time of collection and location of the patient noted on the specimen. Incorrectly labelled specimens will normally be discarded. The container should be firmly closed and carefully sealed in the sealable pocket of the specimen bag with the request card in the outer non-sealable pocket.

Health and safety

The laboratory should be alerted to the potential infection risks from any patient sample by inclusion of sufficient clinical information. High risk stickers should be placed on the samples of patients suspected of having a hazard group 3 or 4 organism. Samples suspected of containing hazard group 4 organisms should not be sent to the laboratory without prior discussion with a consultant Microbiologist.

Important information regarding MRSA screens

MRSA screens are cultured on selective media on which other potential pathogens will not grow. Therefore if there are signs of infection at a wound site, you MUST send a wound swab for full culture, not just as a MRSA screen.

MRSA screens will NOT be processed unless they are ordered on Sunquest ICE.

For more details on MRSA screening refer to the Trust MRSA Policy

Specimen collection

Whenever possible specimens should be collected prior to antimicrobial therapy and promptly delivered to the laboratory.

The laboratory cannot process specimens or interpret the result accurately without suitable and sufficient clinical information. The quality and integrity of the sample is the responsibility of the person collecting the sample. The sample may deteriorate if there is any delay in delivery to the laboratory or if the sample has not been stored incorrectly before delivery.

The laboratory will endeavour to provide the best possible service but the quality of the results ultimately depends on the quality of the sample submitted for testing.

Histopathology and Diagnostic Cytology

Key contacts

Lead Clinician

Dr Samita Agarwal
Direct line 01438 288031 (ext8031)
Secretary 01438 284048 (ext4048)

Consultant Histopathologists:

Dr A Narula
Direct Line 01438 286326 (ext 6326)
Secretary 01438 288040 (ext 8040)

Dr W Mohamid
Direct Line 01438 285279 (ext 5279)
Secretary 01438 286074 (ext 6074)

Dr S Agarwal
Direct Line 01438 288031(ext 8031)
Secretary 01438 284048 (ext 4048)

Dr S Angra
Direct Line 01438 286165(ext 6165)
Secretary 01438 286124(ext 6124)

Dr K Adu-Poku
Direct Line 01438 286181(ext 6181)
Secretary 01438 288042 (ext 8042)

Dr L Mears
Direct Line 01438 284592(ext 4592)
Secretary 01438 288041 (ext 8041)

Dr R Swamy
Direct Line 01438 284158 (ext 4158)
Secretary 01438 285708 (ext 5708)

Dr Y Thevacumar
Direct Line 01438 285282 (ext 5282)
Secretary 01438 288043 (ext 8043)

Laboratory Manager:

Rachel Smith
r.smith96@nhs.net

General Queries & Results:

Office 01438 288043 (ext 8043)
Histology Laboratory 01438 288044 (ext 8044), 01438 285198 (ext 5198)
Cytology Laboratory 01438 285913 (ext 5913)

MDT Coordinators:

Tia Delamore 01438 286182 (ext 6182)
Kelly O'Reilly 01438 286182 (ext 6182)

Opening hours

Monday to Friday: 08.30 – 17.00

The department is situated in the Pathology Department, Level 3 at Lister Hospital, Coreys mill lane, Stevenage SG1 4AB

Specimens and tests

Request Form:

Cellular Pathology requests are now raised using Sunquest ICE and samples labelled with the printed barcodes.

If the ICE system is experiencing downtime the following labelling system must be followed

Addressograph labels with all the patient's identifiable details on will be accepted as well as hand written request forms but they **must** be legible and must have a minimum of **three** patient identifiable features. These are:

- Patients Surname and Forename
- Patients Date of Birth
- NHS number (10-digit number ***-***-****)
- Hospital number
- Patients Address

Request forms should also be filled in legibly with the following details:

- Requesting source/location (i.e. GP surgery, ward, out-patients, etc)
- Requesting Consultant or GP
- Nature of the specimen/site of specimen
- Signature
- Bleep/Contact number
- Brief clinical details

This information is vital to the correct interpretation of the test material and for ensuring that results are sent to the appropriate location. Please also include an extension or bleep number, particularly for urgent samples.

ICE requesting is strongly encouraged however request forms are available from main pathology (ext. 5232).

Histology Samples:

Sample containers should ideally be labelled with the ICE barcode sticker. It *must* be labelled with at least 3 identifiers – Surname, forename, DOB and/or NHS number.

If more than one sample is collected for a patient each sample should be clearly referenced on the request form.

Fixation

Small endoscopic or needle biopsies can be satisfactorily fixed in 2 hours. Larger fragments such as endometrial curettings and colposcopic biopsies need 6 hours fixation or more. Penetration of dense tissue (such as prostatic chippings) or collagen (especially skin) is slow, and these tissues require at least 18 hours fixation. Major resections commonly require overnight fixation to make the tissue firm enough to cut, followed by a second period of overnight fixation to fix the tissue blocks. Inadequate fixation before processing produces poor preservation, which in some cases will make the biopsy impossible to interpret and in many cases will reduce the yield of information.

Note: Formalin contains formaldehyde, which is toxic. All departments must be aware of how to deal with a formalin spillage and have risk assessments and training available in their department to help minimise this risk. MSDS sheets can be obtained from Pathology. Any formalin spillage should be acted upon immediately and recorded as an Enhance.



Small Biopsy Specimens in Formalin

Most biopsies, especially mucosa, become very distorted by shrinkage during fixation, which considerably reduces the information which can be obtained; this can be virtually eliminated by flattening the biopsy gently, mucosal surface up, on a piece of fine card (coarse fibres such as those in blotting paper, paper towel, or gauze can make section cutting impossible); it should be allowed to adhere for about 30 seconds before being placed in formalin. The volume of formalin should be at least ten times the volume of the tissue.

Specimens containing bone:

Bone Marrow Trehines require decalcification and the **minimum** turnaround time for a report is 5 days. **Other specimens**, which contain bone, will take longer than 1 week.

Product of Conception

Product of conception specimens including specimens from suspected ectopic pregnancy specimens must always be accompanied by a **completed consent form**.

Frozen Sections

All frozen sections should be booked in advance if possible. Inform the laboratory of the date and the approximate time of the operation, name of the patient, type of specimen and the reasons for the frozen sections and if the patient is under general or local anaesthetic. The specimen should **NOT** be put in formalin but should be sent to the laboratory immediately. The reporting pathologist will telephone a result within 20 minutes.

Larger Specimens in Formalin

The specimen should be sent in a sealed container of sufficient size to permit easy removal.

Specimens up to about 50ml can be sent in a disposable 350ml container, filled with formalin.

Specimens larger than this should be placed in a plastic bucket of appropriate size and covered with formalin.

The container should be sent to the Department within 2 hours and before 4.00pm, so that it can be opened/sliced and immersed in adequate formalin. Delay produces unsatisfactory fixation and potential loss of important information such as tumour type, depth of invasion, and margins.

We recommend that resected organs are not opened in theatre by the Surgeons themselves, but that specimens are transferred as soon as possible to the Histopathology Department. However, if Surgeons feel that they need to open organs, we suggest the following guidelines:

Segments of intestine should first be opened along the anti-mesenteric border, and stomach along the greater curvature.

The uterus should be opened by transverse section through the endocervical canal 2cm above the external os, followed by a single median slice through the anterior wall down to endometrium.

Other organs should not be opened or sliced, as the probability of information being lost by unskilled slicing is greater than the probability of loss by poor fixation. A delay of several hours in providing adequate exposure to adequate volumes of fixative will inevitably cause poor preservation and should be avoided if at all possible.

Special Fixatives

Specimens for immunofluorescence: a special transport medium (Michel's media) is available from St John's Institute for Dermatology, London. Specimens must be dispatched direct to St Johns **NOT** to the Histopathology department. Tissue left dry or placed in formalin is unusable.

Processing

The standard processing schedule is overnight, from 5.00pm to 8.00am, and tissue must therefore be adequately fixed by about 4.00pm to catch the schedule. The tissue blocks are embedded and sections are cut and stained the following day.

Reporting of Results

Urgent small biopsy results can be available within 48 hours in exceptional cases telephone escalation to Manager. Routine and complex specimens will take longer.

- Total 7 day TAT: 90% (cases for MDT)
- Total 10 day TAT: 80% (all cases)
- Total 21 day TAT: 95% (all cases)

Report distribution

Reports are entered on the computer, authorised and printed; the printed reports are available on the Sunquest ICE 15 minutes after authorisation. Printed reports are dispatched twice per day to both wards and GP surgeries.

Frozen section reports are telephoned to requesting surgeon after reporting. Please ensure you provide a contact number on the request form.

Private reports after authorisation are emailed and/or posted as required.

Lab activities

- Examination of tissues in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis
- Tissue processing
- De-calcification
- Tissue embedding
- Microtomy
- Cryotomy
- Routine morphological staining for the detection of Basophilic and eosinophilic structures
- Special stains for the detection of:
 - Acid mucopolysaccharides
 - Acid and Neutral Mucopolysaccharides
 - Helicobacter Pylori
 - Neutral Mucopolysaccharides, glycogen and fungus
 - Elastic fibres and connective tissue
 - Helicobacter/Microorganisms
 - Gram positive and negative micro-organisms
 - Fungi (Aspergillus) and Pneumocystis
 - Elastic fibres and connective tissue
 - Melanin, Argentaffin cells and Lipofuscin pigment
 - Melanin
 - Connective tissue and fibrin
 - Copper assoc. Protein and Hepatitis B
 - Neutral Mucopolysaccharides, glycogen and fungus
 - Reticulin fibres
 - Mast cells
 - Calcium
 - Tubercl bacilli
- Immunohistochemistry to detect the following:
 - Smooth muscle cells, myofibroblasts, myoepithelial cells, Diagnosis of leiomyomas, leiomyosarcomas
 - Broad band cytokeratin marker, epithelial cell carcinomas
 - HGPIN, prostate adenocarcinoma
 - BCL-2 oncoprotein
 - BCL-6 gene
 - Epithelial cell types
 - Ovarian cancer
 - Carcinomas, RCC
 - Reacts with human calretinin and intracellular calcium binding protein. Marker for mesotheliomas
 - T cells
 - Mature T cells, T cell lymphomas
 - Granulocytes Reed-Sternberg cells
 - Mature B cells, Follicular dendritic cells
 - B cells, some T cells
 - B Cells, Reed-Sternberg cells
 - Reed-Sternberg cells and ALCL
 - Haematopoietic cells, vascular endothelium
 - T cells and some soft tissue

- Leukocyte common antigen
- Neuroendocrine cells
- Macrophages
- B lymphocytes
- GIST
- Plasma cells
- Adenocarcinoma and carcinoid (intestinal epithelium)
- CEA glycoproteins in adenocarcinoma
- Neuroendocrine tumours
- Stratified squamous epithelium, basal cells, mesotheliomas
- Normal and neoplastic epithelia
- Glandular and transitional epithelium
- Basal cell carcinoma, squamous cell carcinoma
- Adenocarcinomas
- Adenocarcinomas, columnar epithelial cells
- Striated and smooth muscle cells
- Normal duct epithelial cells, ductal carcinoma
- Nuclei of cells containing a high level of oestrogen
- V.W factor in endothelial cells
- Mesothelial Cells Thyroid carcinomas
- Melanoma, melanocyte differentiation
- Proliferating Cells
- B cells and plasma cells
- Melanoma marker
- Epithelial tissue from glandular to stratified squamous
- Peripheral nerves, neuroendocrine tumours
- Seminomas
- HPV driven tumour e.g. Squamous cell carcinoma
- Neoplastic cells in epithelium
- Squamous epithelium
- Nuclei of cells showing expression of Progesterone
- Prostate secretory and ductal epithelium
- Schwann cells, nerve processes, S100 +ve neoplasms eg Melanoma
- Smooth muscle cells, myoepithelial cells
- Neuroendocrine cells
- TTF-1 found in lung and thyroid (thyroid follicular cells)
- Cells of mesenchymal origin
- Epithelial cells and smooth muscle in fallopian tube. Wilms tumour
- Human Epidermal Growth Factor Receptor 2 (HER 2) oncoprotein expression in breast cancer

Non-Gynae Cytology activities

- Fine needle aspirations. (FNA) of easily accessible solid or cystic lesions. e.g. breast, thyroid, lymph nodes, salivary glands, subcutaneous masses.
- Respiratory samples eg. Sputum, bronchial washings
- Urine
- Body cavity fluids e.g. ascitic, pleural, synovial.
- Brushings eg. CBD, nasal, gastric, oesophageal
- CSF
- Skin scrapes
- Joint fluid-crystal examination

Please note. Cells degenerate rapidly. Samples for cytological examination must be sent to the laboratory as soon as possible. Enquiries regarding specimen collection should be directed to the laboratory on ext 5913.

Collection/transport fluid, Cytolyt® is available from the Cytology Dept.

Pathologists are available to undertake skin scrapes and Fine Needle Aspirations (FNA) on palpable masses. Please contact the office 01438 288043 or ext. 8043 for further information.

Gynae Cytology send samples to NNUH via the purple bag system.

Fine Needle Aspirates (FNA)

- Make 1-2 air-dried slides & wash needle in a collection fluid, Cytolyt® solution, supplied by the Cytology department (or sterile saline can be used if necessary)
- Do NOT reuse needle to repeat FNA, use a new needle.
- Label slides with a pencil, place in the transport box, label Universal container of washings and send to the laboratory.
- Complete request form including patient details, specimen type and site, and appropriate clinical information - including date of next clinic appointment if appropriate, and destination that results are to be sent to.
- Send sample and request form to the laboratory immediately.
- For Thyroid FNA take 6 slides in total, 3 air-dried and 3 fixed with Cytofix provided by the Cytology Department.

Respiratory samples

- Sputum - a sputum pot of an early morning deep cough specimen, should be sent on 3 successive days.
- Bronchial washings - sample container of wash/trap specimen without fixative.
- Bronchial brush - place cut end of brush into collection fluid (Cytolyt® solution).
- Do NOT leave brush inside the sheath.

Urine

- A Universal container of freshly voided urine - **NOT** the first specimen of the day.
- If not a voided specimen please identify the type e.g. catheter, ileal conduit or cystoscopy
- Send immediately to laboratory.

Fluids

- Send a fresh specimen.
- Do NOT send entire bag - Disperse any sediment and send an aliquot of fluid in 1-2 Universal containers.
- Peritoneal and other washouts should be clearly identified as such.

Brushings

- Cut off the end of endoscopy brush and place in a cytology collection fluid (Cytolyt® solution)
- Please identify the site and give relevant clinical details on the request form.
- Send to the laboratory as soon as possible.

CSF

- Cytology is for the detection of abnormal cells.
- Please send to the laboratory immediately.
- Requests for all cell counts for different departments will require separate samples and ICE requests, ie for Micro/ PHE.

Skin scrapes - This is done by Dr R Swamy Booking follow the frozen section path

- These should be spread thinly and evenly across a labelled glass slide.
- If possible, 2 slides – one fixed immediately without drying, the other air-dried rapidly.

Joint fluid-crystal examination

- Fresh sample in plain universal container

Turnaround times for diagnostic specimens

The Cytology Department aims to provide a quality service to its users and prompt turnaround times are essential in achieving that aim. The following turnaround times may be used as a guide:

Urgent specimens: 24 hours - Urgent specimens must be clearly marked as **Urgent** and a contact telephone or bleep number must be given on the request form.

Non-urgent specimens: 72 hours - *Please Note:* For any specimen, the Consultant Pathologist may request the laboratory to carry out additional ancillary procedures to assist with the diagnosis. This may delay the issue of a final report. In these cases an interim report either written or verbal may be issued by the reporting pathologist.

Referral of Tests in Cellular Pathology

The Cellular Pathology department based at The Lister Hospital has a large repertoire of special staining techniques and an extensive panel of immunohistochemistry test in-house.

On occasions it may be required that specific tests not performed in-house are sent elsewhere to referral labs. Please refer to table below for list of referral tests which are sourced externally:

| Laboratory Name: | Address: | Referral Tests performed: | TAT for referral Test |
|----------------------------|---|---|---|
| HSL-AD | HSL-Advanced Diagnostics Ground Floor 60 Whitfield Street London W1T 4EU | IHC tests which are not performed routinely in-house ALK, ROS, PDL-1 All FISH assays (inc HER2, ALK, ROS) | 24 hrs 5- 8 working days (includes interpretation) 5 Days* |
| Sarah Cannon Molecular Lab | Sarah Cannon Molecular Lab Shropshire House 1 Capper Street Fitzrovia London WC1E 6JA | NGS (Multigene Panel) MLH-1 Hypermethylation BRAF single gene EGFR single gene (rapid) | 8-10 working days 5 working days 5-7 working days 48 hrs |
| Source Bioscience | Source Biosciences 1 Orchard Place Nottingham Business Park Nottingham NG8 6PX | Gastric HER2 | 5 days |
| North Thames Genomic Hub | Clinical Genomics MolecularDiagnostics Dept The Centre for Molecular Pathology The Royal Marsden NHS Foundation Trust 15 Cotwold Road Sutton, Surrey SM2 5NG | NTRK fusion testing | 10-14 days |

Cervical Cytology Samples

Cervical cytology service is provided by Norfolk and Norwich Laboratory at the following address.

Cervical Cytology Laboratory

Norfolk and Norwich University Hospital
Colney Lane
Norwich
NR4 7UY
Contact: 01603 287412

Email: nnu-tr.cytology@nhs.net

Sample preparation

Sample vials *must* be labelled with at least 3 identifiers – Surname, forename, DOB and/or NHS number.

All routine cervical screening samples should be accompanied by the national request form HMR 101/5.

The patient's full name, address, DOB and NHS number *must* be written on all request forms and all parts of the form should be completed.

Users are encouraged to use the pre-printed HMR 101 form from the Open Exeter system

If you have any difficulty in accessing Open Exeter, contact Andrew Martin, NHS Hertfordshire, 01707 369756.

Cervical samples from hospital clinics should be accompanied by the white and purple three-part Histology Request form.

Turnaround times for cervical cytology

From December 2010 all cervical screening services have to ensure a 14-day turnaround time. This is from the date of the sample being taken to the time that the woman receives her result letter. It is essential that all cervical samples are sent to the laboratory immediately in order to help achieve this target.

Further information can be found in the NHS CSP document: Cytology improvement guide – achieving a 14 day turnaround time in cytology. (November 2009). Please see the link below.

<http://www.cancerscreening.nhs.uk/cervical/14-tat.html>

Cervical Cytology Sample Taker Training

For information on sample-taker training please contact Roseanna Bignell, Network Laboratory Manager on the telephone number or email address shown above.

Annexure A: Referral Laboratories

| Code | Full address |
|------|---|
| ADI | Department of Clinical Biochemistry & Clinical Immunology Box 232, Level 4, Addenbrookes NHS Trust Hills Road, Cambridge, CB2 2QQ Tel: 01223 336792 |
| AGH | Cambridge Genomic Laboratory, Box 143, ATC level 6 Cambridge University Hospital Foundation Trust, Addenbrooke's Hospital Hills Road, Cambridge, CB2 0QQ |
| AML | Antimicrobial Reference Laboratory Level 2, Phase 1, Pathology Sciences Building Southmead Hospital Westbury-on-Trym Bristol, BS10 5NB Tel: 0117 4146220 |
| BED | Clinical Biochemistry Bedford Hospital NHS Trust Kempston Road, Bedford, MK 9DJ Tel: 0 1234795915 |
| CHX | The SAS Laboratories Clinical Biochemistry & Medical Oncology Charing Cross Hospital Fulham Palace Road, London, W6 8RF Tel: 020 3313 5353 |
| CAR | Cardiff Toxicology Laboratory 4 th Floor, Academic Centre University Hospital Llandough Penlan Road, Llandough, Penarth Vale of Glamorgan, CF64 2XX Tel: 029 2071 6894 |
| CVU | Cardiff and Vale University Health Board UHW University Hospital of Wales Heath Park Cardiff CF14 4XW Tel: 029 2074 7747 |
| EKE | Immunology Laboratory William Harvey Hospital Kennington Road Willesborough Ashford Kent TN24 0LZ Tel: 01233 616287 |
| GOS | Chemical Pathology Great Ormond Street Hospital for Children Great Ormond Street, London, WC1N 3JH Tel: 0207 405 9200 and dial 5009 |
| HALO | Health Services Laboratories The Halo Building 1 Mabledon Place London, WC1H 9AX |
| HOM | Dept of Clinical Chemistry Homerton University Hospital NHS Foundation Trust Homerton Row, Hackney, E9 6SR Tel: 0208 510 7887/7888 |
| ION | Department of Neuroimmunology & CSF Laboratory Room 917, Institute of Neurology Queen's Square, London, WC1N 3BG Tel: 0203 4483814/3844 |
| RDE | Synnovis Analytics LLP Department of Clinical Biochemistry Kings College Hospital Denmark Hill, London, SE5 9RS Tel: 020 3299 4126 |
| KIT | Toxicology Laboratory 3rd Floor, Bessemer Wing King's College Hospital NHS Foundation Trust |

| | |
|-----|--|
| | Bessemer Road, Denmark Hill London SE5 9RS Tel: 020 3299 5883 |
| LTH | The Department of Blood Sciences, Leeds Teaching Hospitals NHS Trust, The Old Medical School, Great George Street, Leeds, LS1 3EX |
| MRI | Clinical Biochemistry Manchester Royal Infirmary Oxford Road Manchester M13 9WL Tel: 0161 276 8766 |
| MTL | Central Specimen Reception North Wing - 5th Floor St Thomas' Hospital Westminster Bridge Road London SE1 7EH |
| MWY | Biochemistry Department Clinical Science Building Wythenshawe Hospital Manchester University NHS Foundation Trust Southmoor Road Manchester M23 9LT Tel: 0161 291 2126 . |
| NNH | Norfolk and Norwich University Hospitals NHS Foundation Trust Calcium and Bone Metabolism Laboratory Laboratory Medicine Level 1, East Block Colney Lane Norwich NR4 7UY |
| NOT | Molecular Diagnostics Section Dept Clinical Pathology Nottingham University Hospitals Queens Medical Centre Campus Nottingham NG7 2UH |
| OXF | Clinical Laboratory Immunology Churchill Hospital Churchill Drive Old Road Headington Oxford OX3 7LE |
| QUE | Neurometabolic Unit (Box 105) 6th floor, Institute of Neurology Queen Square House Queen Square London WC1N 3BG Tel: 020 344 83818 |
| RSC | SAS peptide Hormone Section, Clinical Laboratory Royal Surrey County Hospital Egerton Road, Guildford, GU2 5XX Tel: 01483 406715 |
| STS | Supra-Regional Assay Laboratory Specialised laboratory Medicine DU [Chemical Pathology] 5 th Floor, North Wing, St Thomas Hospital Lambeth Palace Road, London, SE1 7EH Tel: 020 7188 7188 |
| SAN | Department of Clinical Biochemistry City Hospital Dudley Road Birmingham |

| | |
|------|--|
| | B18 7QH Tel: 0121 507 3517 |
| SGP | Protein reference Unit & Immunopathology Level 2, Jenner Wing, St George's Hospital Tooting, London, SW17 ONH Tel: 0208 725 0025 |
| SIH | UCLH SIHMDs SIHMDs Flow Cytometry Level 2 Halo Health Services Laboratories The Halo Building 1 Mabledon Place London, WC1H 9AX |
| STH | Department of Chemical Pathology & Metabolism St Helier Hospital Wrythe lane, Carshalton, SM5 1AA Tel: 01372 735258 |
| SHP | Immunology Department & Protein Reference Unit P.O. Box 894 Sheffield, S5 7YT Tel: 0114 2715552 |
| UCLH | University College London Hospital, Special Chemistry 3 rd Floor, 60 Whitfield Street London, W1T 4EU Tel: 020 3447 9405 |
| QEH | University Hospitals Birmingham NHS Foundation Trust Clinical Laboratory Services, Level -1 Queen Elizabeth Hospital Mindelsohn Way Edgbaston Birmingham B15 2WB |

Annexure B: Sunquest ICE Order Communication System

Requesting Pathology tests and accessing results using Sunquest ICE

We strongly recommend interaction with Sunquest ICE software for requesting pathology and accessing results for the following disciplines:

- Blood Sciences (Haematology, Clinical Biochemistry, Immunology)
- Cytology
- Microbiology
- BT (additional labelling criteria see BT specific section if Handbook)
- Cellular Pathology

Instructions for the use of Sunquest ICE are available from the Pathology section on the Knowledge centre (Clinical departments, Pathology, Pathology handbook - Pathology User Guide / ICE requesting tests / ICE reviewing results).

Training for ICE - user accounts and training enquiries should be directed to the ICE team at enhtr.iceocs@nhs.net or on Lister extension 4798.

Specimen labelling for Sunquest ICE requests

All specimens must be accurately labelled with at least the minimum patient identification details listed in section 4. The specimen should be labelled at the time of collection. All specimens should be labelled with the ICE OCS generated bar-code label or if necessary, completed by hand. Use of addressograph labels on specimen containers, other than Histology, is forbidden. (This has been shown to be linked with increases in sample mislabelling).

Samples for blood transfusion must be hand-written. ICE OCS generated bar-code labels are not acceptable for transfusion purposes.

Annexure C: Request Form



HEALTH SERVICES
LABORATORIES



Hertfordshire and
West Essex Integrated
Care System

LABORATORY USE ONLY

PATIENT LABEL HERE

Hertfordshire and West Essex ICS

Pathology Request Form

ADDRESSOGRAPH LABEL / PATIENT DETAILS (PLEASE USE BLOCK CAPITALS)

| | | | | |
|----------|--|---------------------|--------------|---------|
| Title | Gender <input type="radio"/> F / <input type="radio"/> M | Different at Birth? | DOB DD/MM/YY | NHS No. |
| Surname | | | | |
| Forename | | | | |
| Address | | | | |
| Postcode | Telephone | | | |

SAMPLE / REQUEST DETAILS

| | | | | |
|--------------------------------|-------------------|-----------|----------|-----|
| Sample Type | Clinical Details | | | |
| Ward / Clinic Name & Address | | | | |
| Request By? | Contact Telephone | | | |
| Sample Date DD/MM/YY | Sample Time HH:MM | Priority? | URGENT | YES |
| Sample Taker's Name / Initials | | ROUTINE | Fasting? | NO |

| BIOCHEMISTRY | |
|---|---|
| <input type="checkbox"/> UE + GFR | <input type="checkbox"/> LIPID PROFILE |
| <input type="checkbox"/> LFT + GGT | <input type="checkbox"/> COAGULATION SCREEN |
| <input type="checkbox"/> CALCIUM | <input type="checkbox"/> PROTHROMBIN TIME & INR |
| <input type="checkbox"/> CRP | <input type="checkbox"/> GLANDULAR FEVER |
| <input type="checkbox"/> GLUCOSE | <input type="checkbox"/> Hb ELECTROPHORESIS |
| <input type="checkbox"/> HBA1C | <input type="checkbox"/> ANF |
| <input type="checkbox"/> TFT (T4/T3) | <input type="checkbox"/> SERUM ELECTROPHORESIS |
| <input type="checkbox"/> FERRITIN | <input type="checkbox"/> THYROID ANTIBODIES |
| <input type="checkbox"/> B12 / Folate | <input type="checkbox"/> COELIAC |
| <input type="checkbox"/> PSA | |
| <input type="checkbox"/> FSH | |
| <input type="checkbox"/> LH | |
| <input type="checkbox"/> PROLACTIN | |
| <input type="checkbox"/> PROGESTERONE | |
| <input type="checkbox"/> TESTOSTERONE | |
| <input type="checkbox"/> URINE MICROALBUMIN | |

| HAEM & IMMUNOLOGY | |
|---|---|
| <input type="checkbox"/> FULL BLOOD COUNT | <input type="checkbox"/> ENT SWAB |
| <input type="checkbox"/> ESR | <input type="checkbox"/> GENITAL SWAB |
| <input type="checkbox"/> COAGULATION SCREEN | <input type="checkbox"/> WOUND SWAB |
| <input type="checkbox"/> PROTHROMBIN TIME & INR | <input type="checkbox"/> URINE MICROSCOPY / CULTURE |
| <input type="checkbox"/> GLANDULAR FEVER | <input type="checkbox"/> C.DIFF |
| <input type="checkbox"/> Hb ELECTROPHORESIS | <input type="checkbox"/> NOROVIRUS SCREEN |
| <input type="checkbox"/> ANF | <input type="checkbox"/> FAECAL PCR |
| <input type="checkbox"/> SERUM ELECTROPHORESIS | |
| <input type="checkbox"/> THYROID ANTIBODIES | |
| <input type="checkbox"/> COELIAC | |

| MICROBIOLOGY & VIROLOGY | |
|---|--|
| <input type="checkbox"/> FULL BLOOD COUNT | <input type="checkbox"/> ENT SWAB |
| <input type="checkbox"/> COAGULATION SCREEN | <input type="checkbox"/> GENITAL SWAB |
| <input type="checkbox"/> PROTHROMBIN TIME & INR | <input type="checkbox"/> WOUND SWAB |
| <input type="checkbox"/> GLANDULAR FEVER | <input type="checkbox"/> URINE MICROSCOPY / CULTURE |
| <input type="checkbox"/> Hb ELECTROPHORESIS | <input type="checkbox"/> C.DIFF |
| <input type="checkbox"/> ANF | <input type="checkbox"/> FAECAL PCR |
| <input type="checkbox"/> SERUM ELECTROPHORESIS | |
| <input type="checkbox"/> THYROID ANTIBODIES | |
| <input type="checkbox"/> COELIAC | |
| | <input type="checkbox"/> FUNGAL MICROSCOPY / CULTURE |
| | <input type="checkbox"/> SPUTUM CULTURE |
| | <input type="checkbox"/> MRSA SCREEN |
| | <input type="checkbox"/> HEP B |
| | <input type="checkbox"/> HEP C ANTIBODY |
| | <input type="checkbox"/> HEP B SURFACE ANTIGEN |
| | <input type="checkbox"/> HIV SEROLOGY |
| | <input type="checkbox"/> SYPHILIS ANTIBODY SCREEN |
| | <input type="checkbox"/> CHLAMYDIA SCREEN |
| | <input type="checkbox"/> COVID-19 SCREEN |
| | <input type="checkbox"/> RESPIRATORY VIRUS PANEL PCR |

OTHER TEST REQUESTS

For Laboratory Use

| SST | EDTA | OTHER | Signature |
|-----|------|-------|-----------|
| | | | |